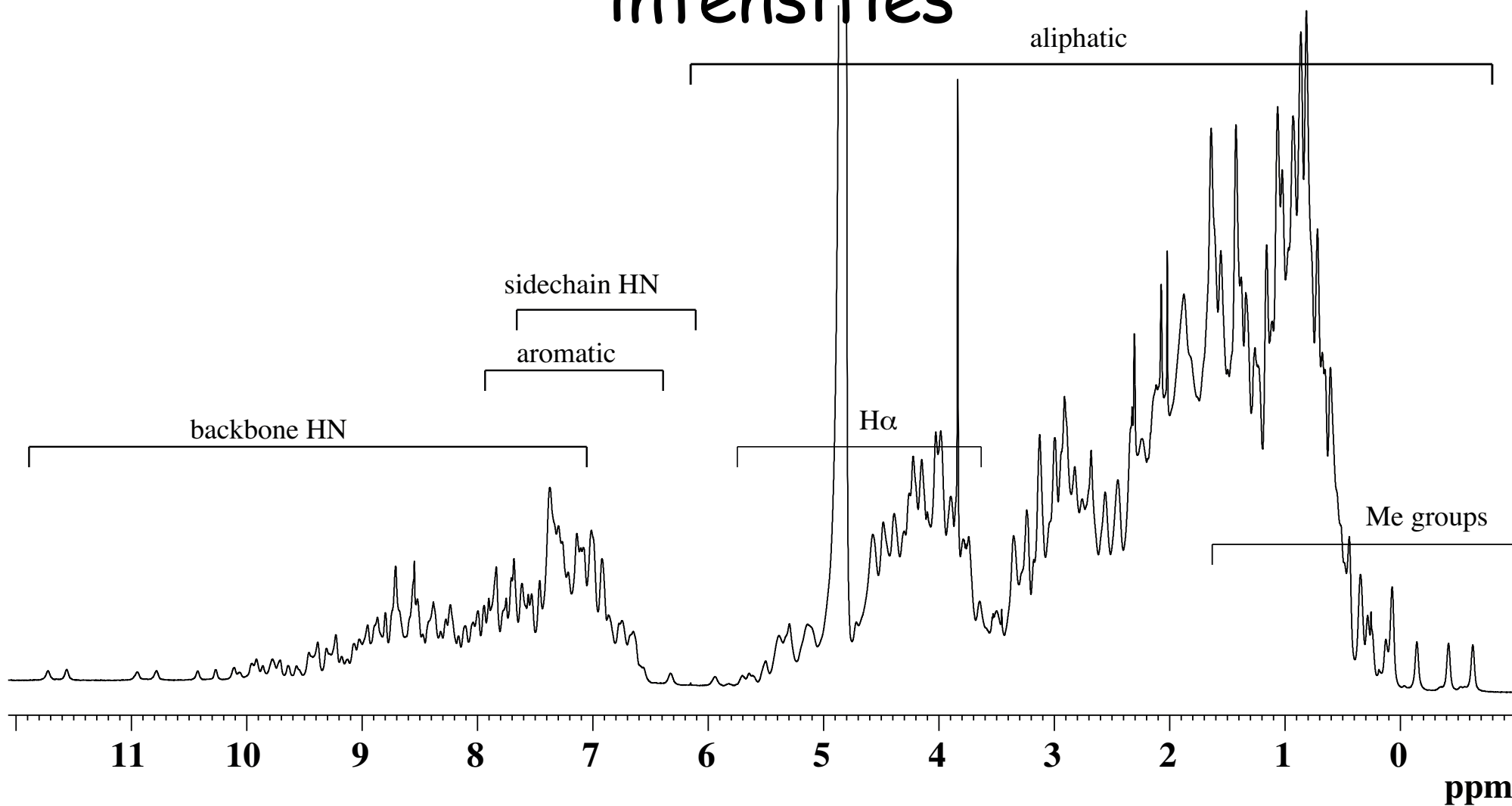
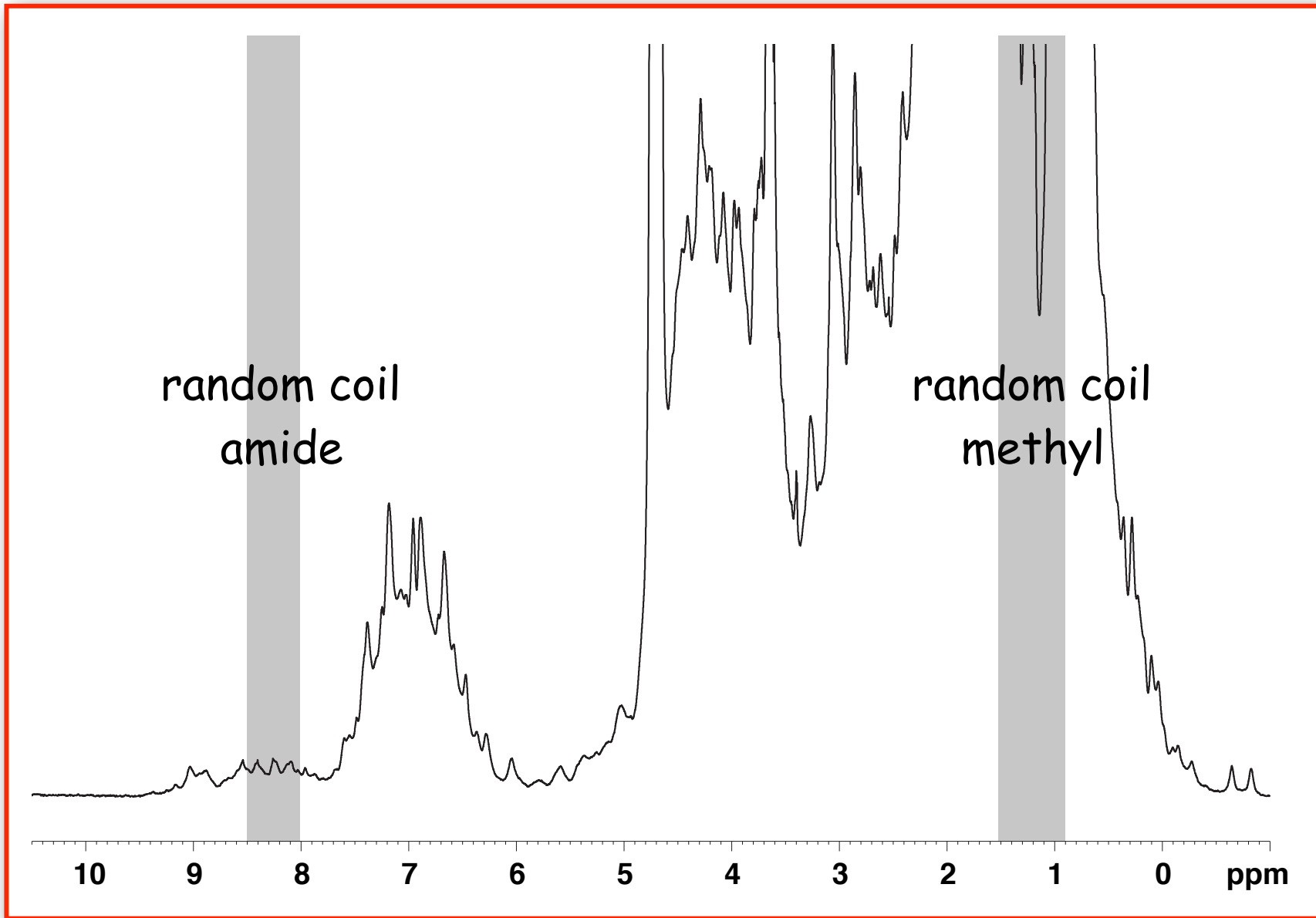


# From NMR spectra to structures

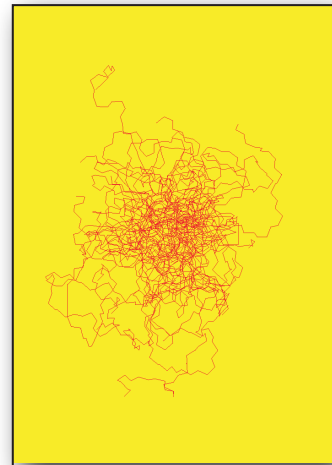
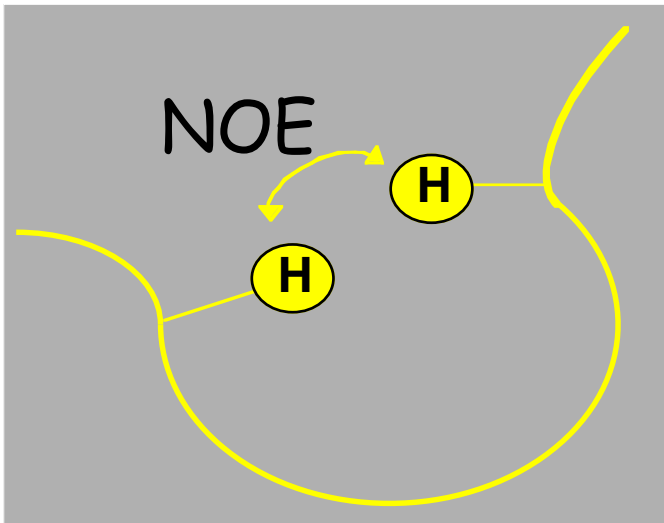
- find conditions under which the protein does not aggregate is reasonably stable
- measure nmr spectra
- sequence-specific sequential resonance assignment
- identify spin systems
- link spin systems (sequential assignment)
- (stereospecific assignment of diasterotopic protons)
- fully- interpret NOESY spectrum
- convert NOESY peak amplitudes into distances
- calculate 3D structure and refine the output

# Chemical shifts (frequencies) and line intensities





NOE information is used to introduce distance restraints into the structure calculations



"NOEs"

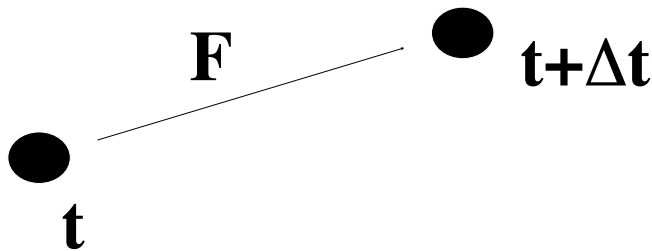


$$\text{NOE} \sim B_{\text{loc}}^2$$

$$B_{\text{loc}} \sim \frac{\gamma_I \gamma_S (3 \cos^2 \phi - 1)}{r^3}$$

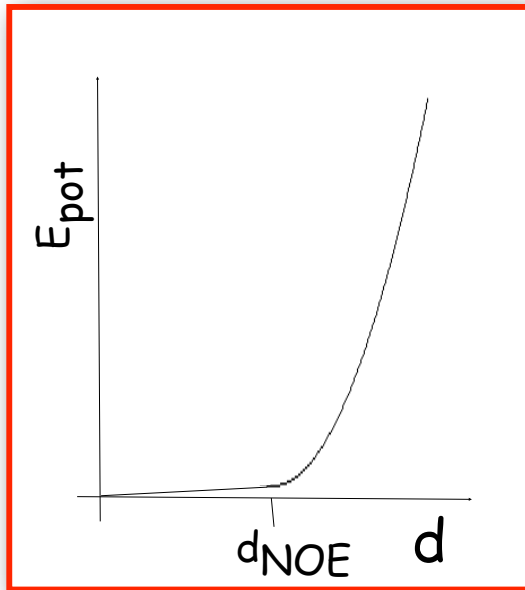
# From distances to 3D structures (II)

- Restrained molecular dynamics:



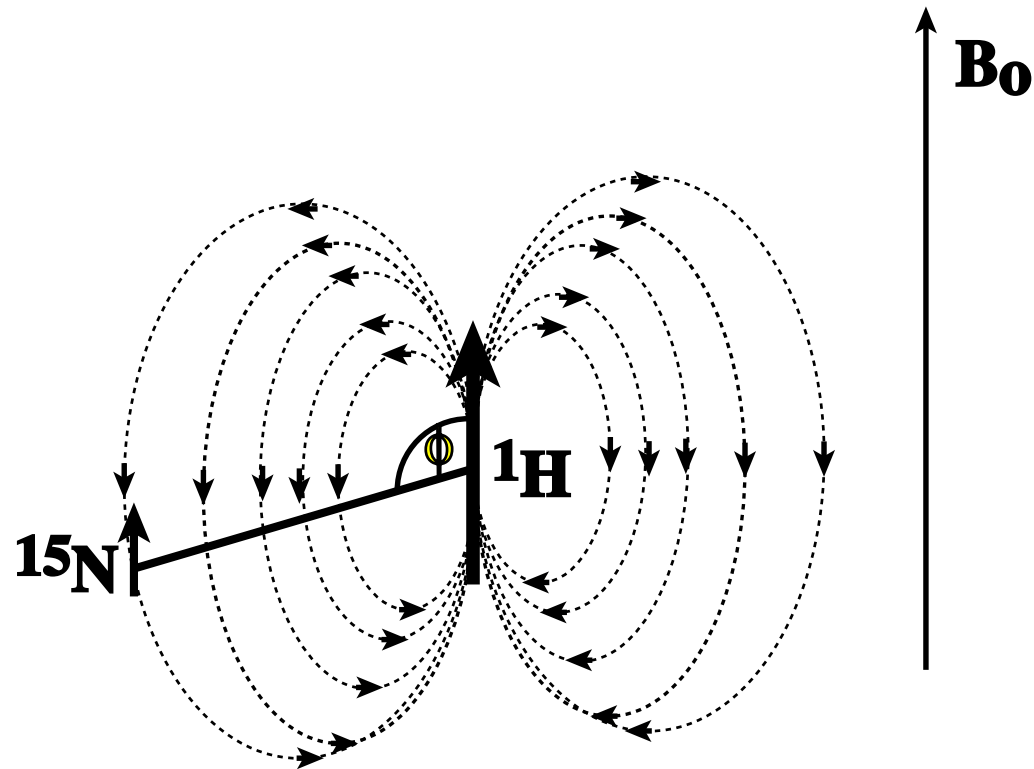
$$F = m \frac{\partial^2 r}{\partial t^2} \quad F = \frac{\partial U_{\text{pot}}}{\partial r}$$

$$U_{\text{pot}} = U_{\text{bond}} + U_{\text{angle}} + U_{\text{dihedral}} + U_{\text{chiral}} + U_{\text{v.d.Waals}} \\ + U_{\text{coulomb}} + U_{\text{NMR}}$$



$$U_{\text{NMR}} = U_{\text{NOE}} + U_{\text{J}} + \dots$$

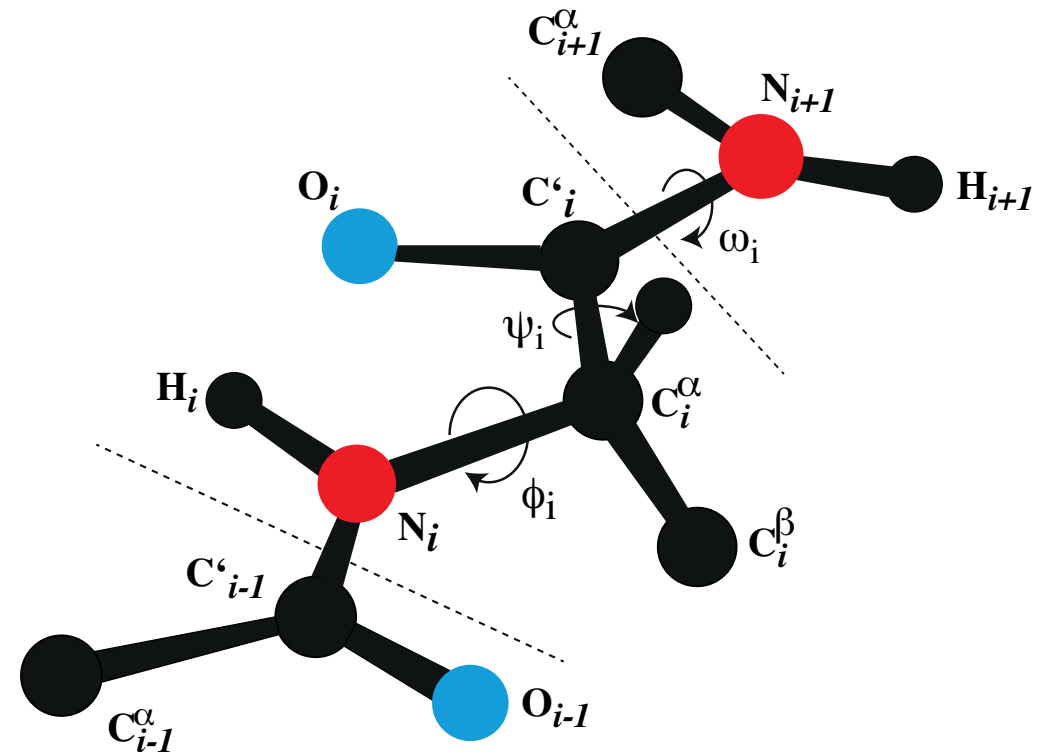
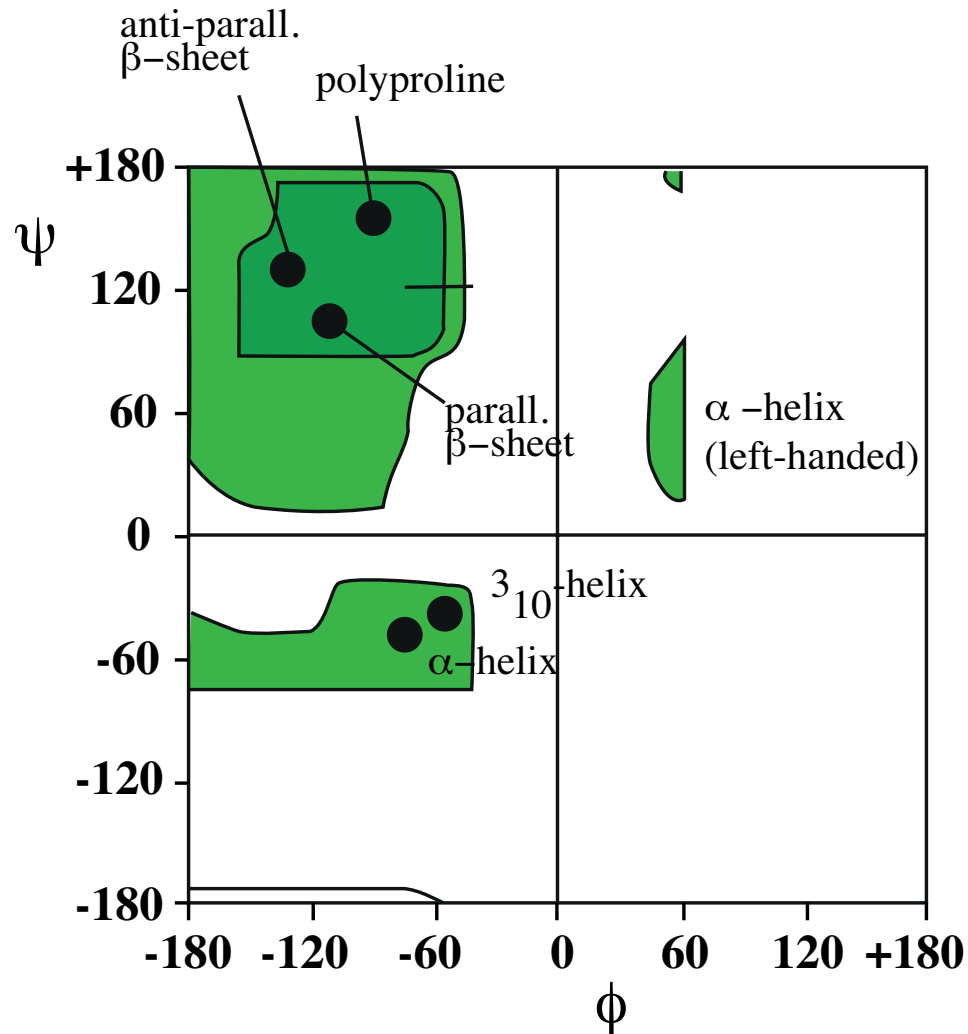
The origin of the NOE is dipolar  
(through-space) coupling of protons

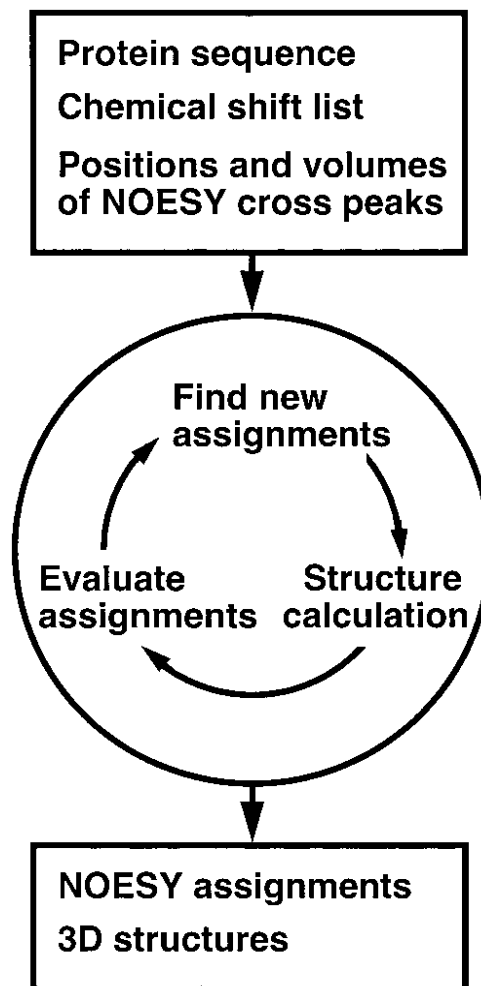


$$B_{\text{loc}} \sim \frac{\gamma_I \gamma_S (3 \cos^2 \phi - 1)}{r^3}$$

$$R_2, R_1 \sim B_{\text{loc}}^2$$

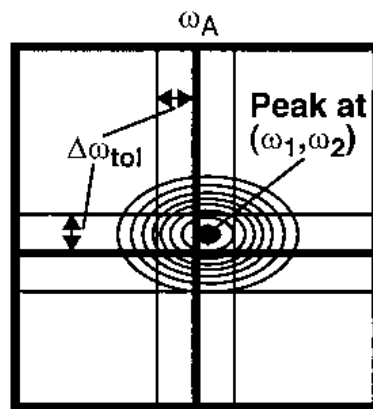
During the structure calculation only rotations about dihedrals are made





# Automated calculation of NMR structures

Chemical shift agreement

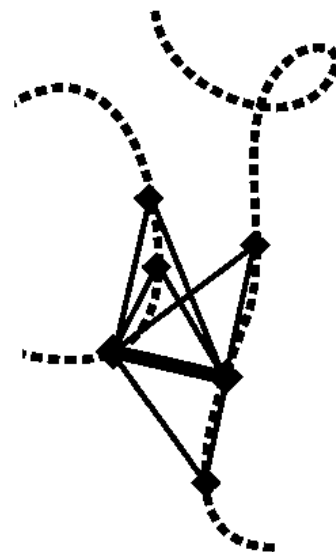


$$|\omega_1 - \omega_A| < \Delta\omega_{tol}$$

$$|\omega_2 - \omega_B| < \Delta\omega_{tol}$$

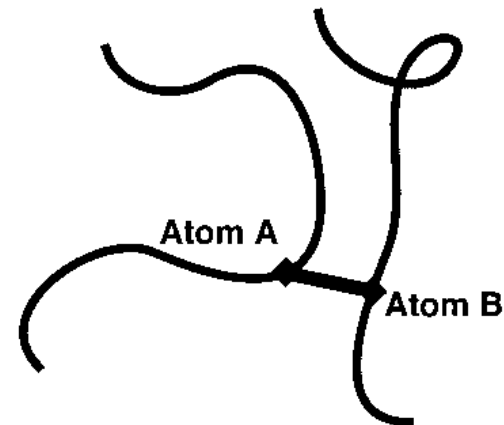
(a)

Network-Anchoring



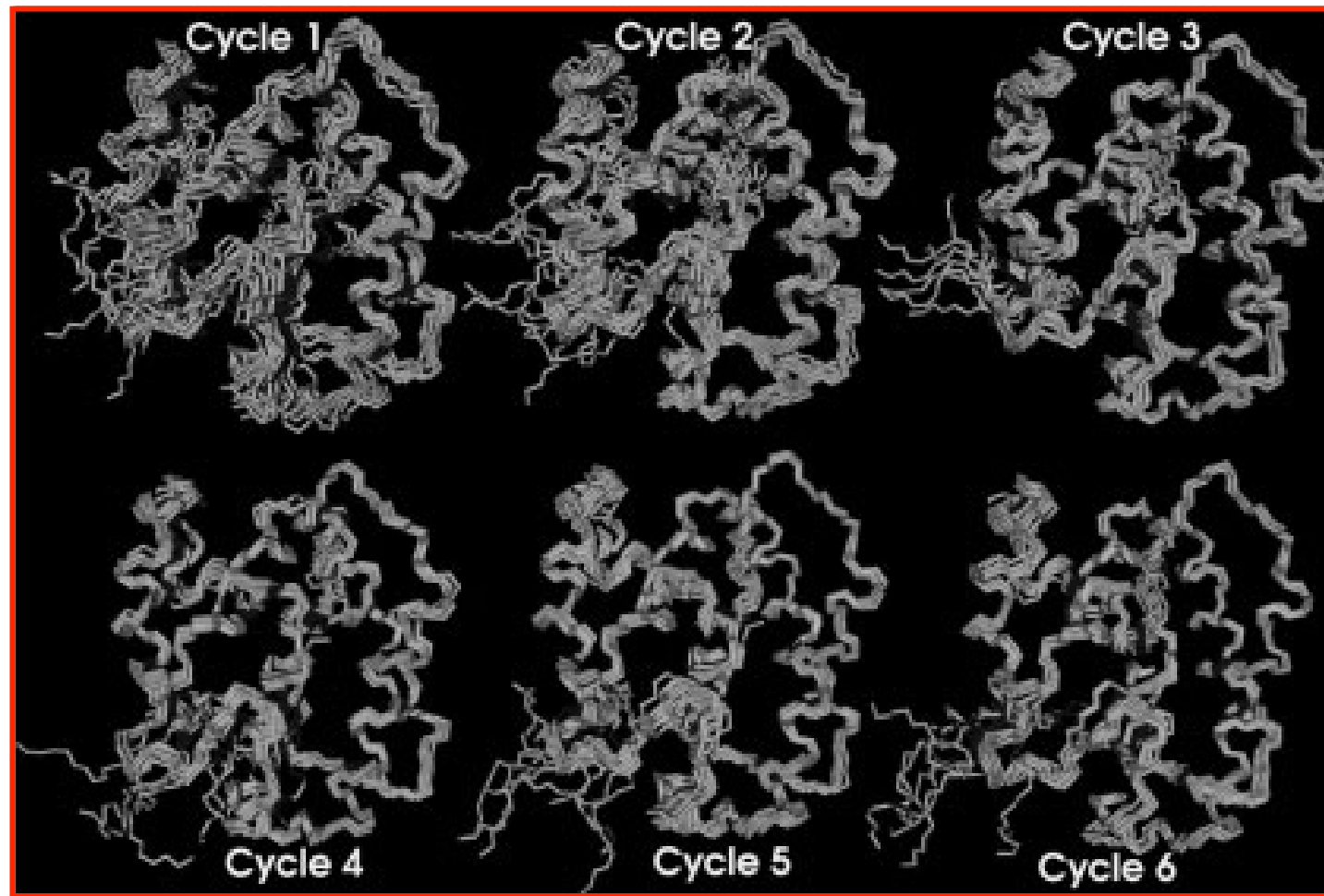
(b)

Consistency with preliminary structure



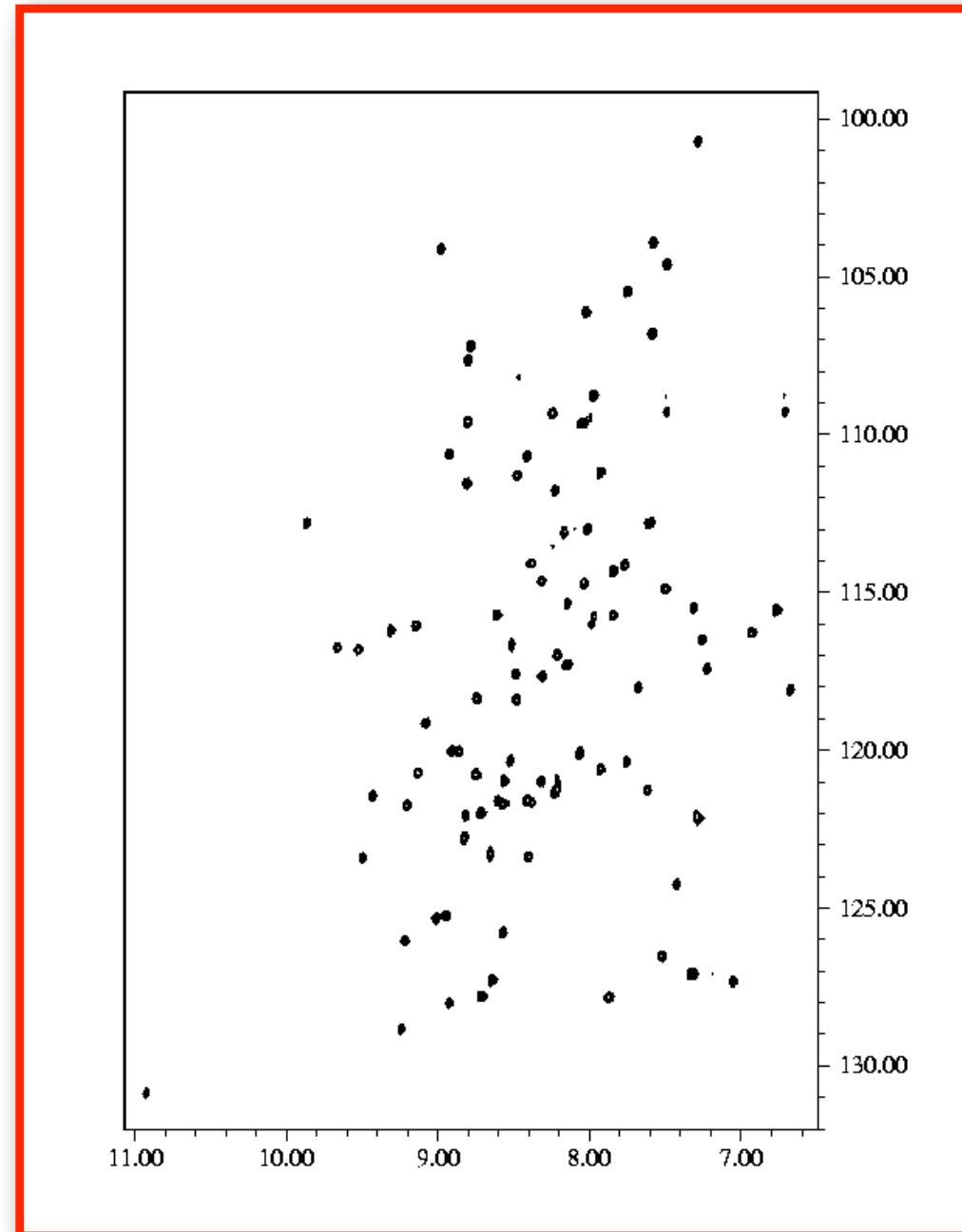
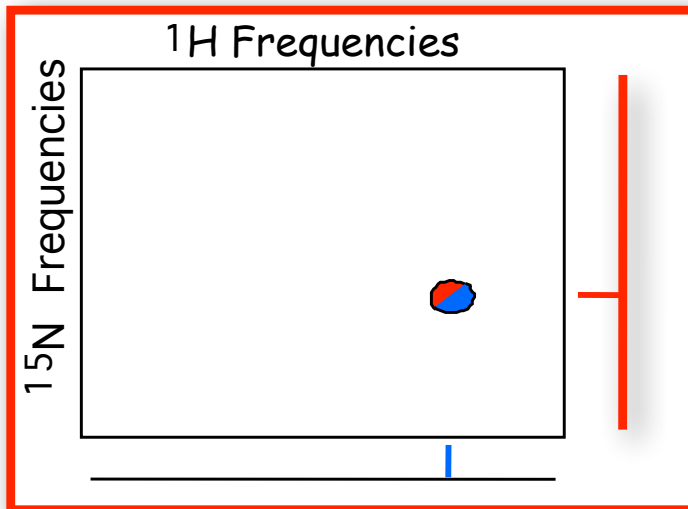
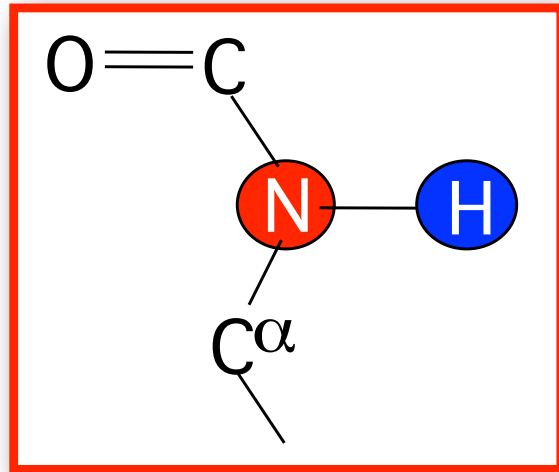
$$d_{AB} < d_{max}$$

(c)

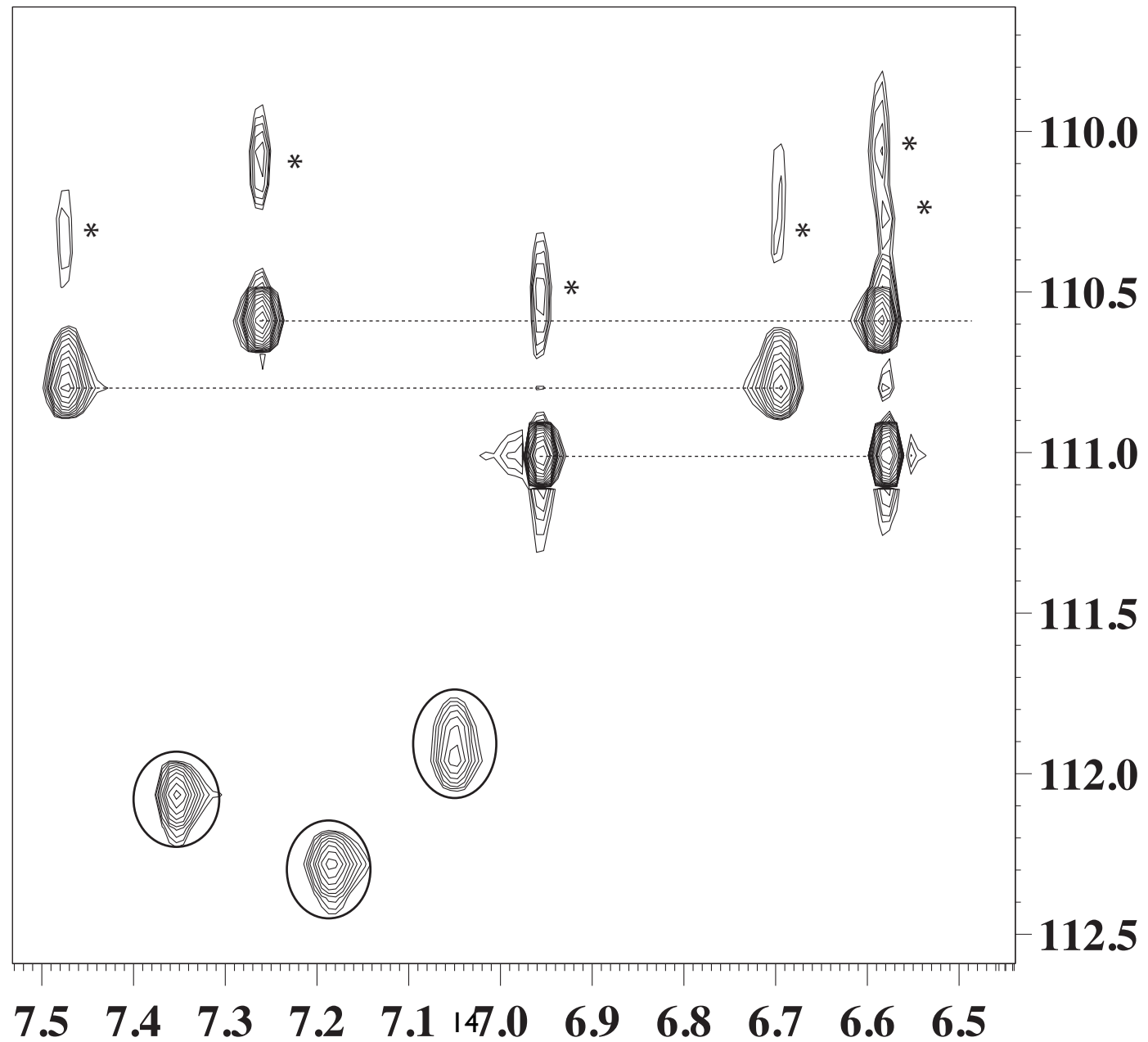


# Methods for assigning larger proteins

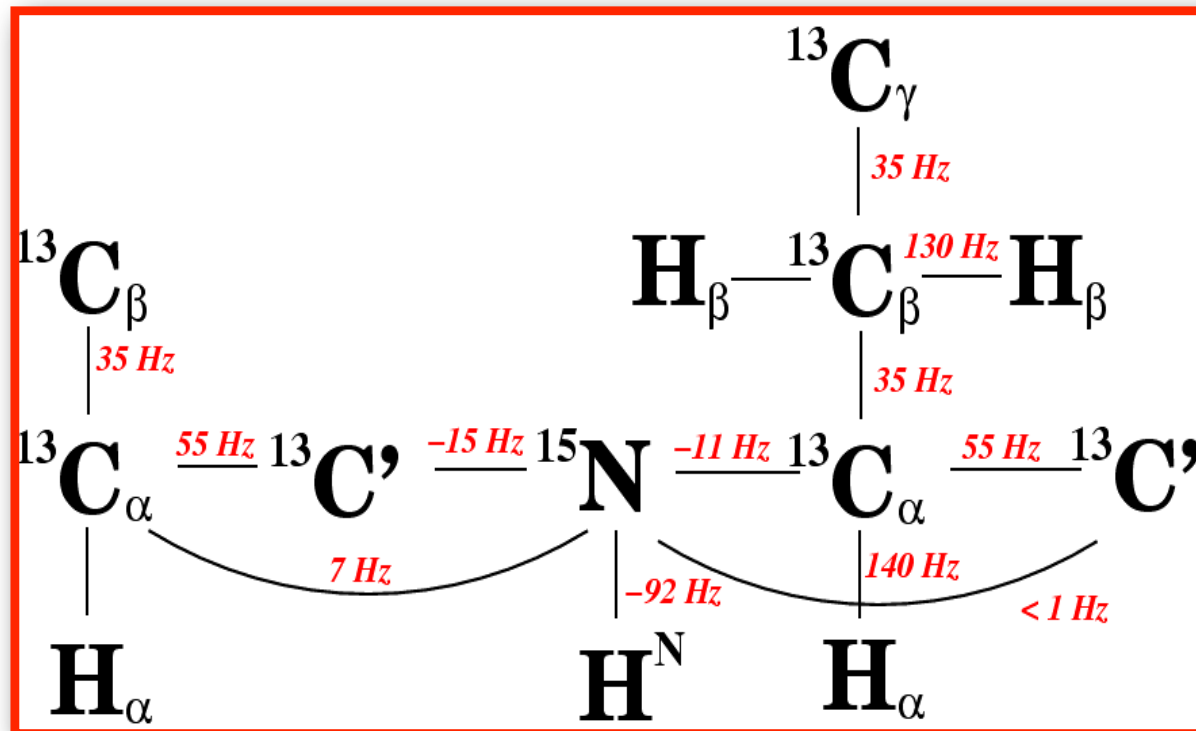
# $^{15}\text{N}, ^1\text{H}$ HSQC spectra are fingerprints of proteins



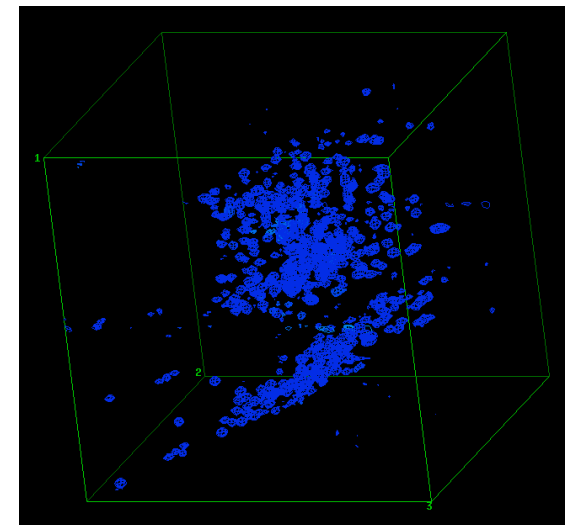
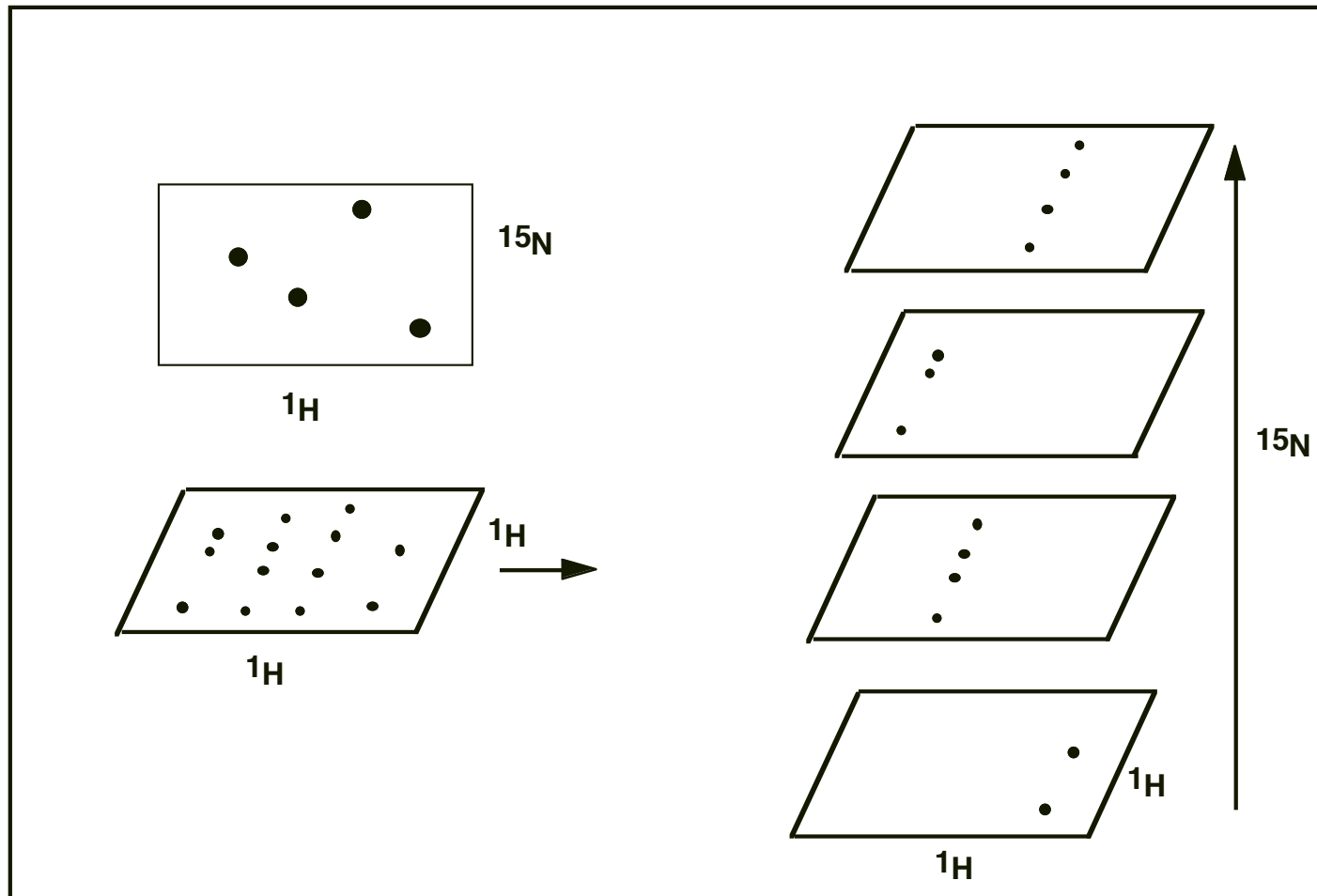
# $^{15}\text{N}$ spectra of proteins: Sidechain peaks



# Sequence-specific resonance assignment in $^{13}\text{C}$ , $^{15}\text{N}$ labelled proteins

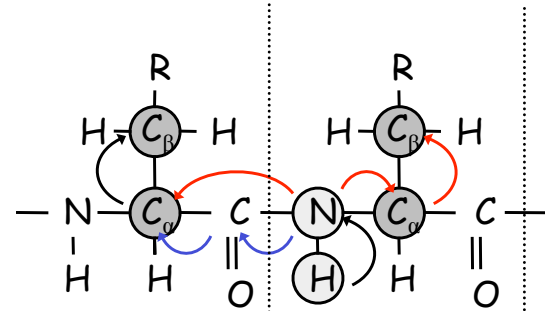


## >> Use of 3-dimensional (tripleresonance) experiments

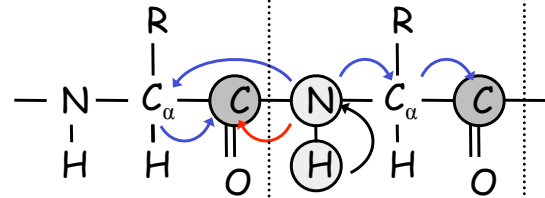


# Backbone Assignment - 3D Experiments

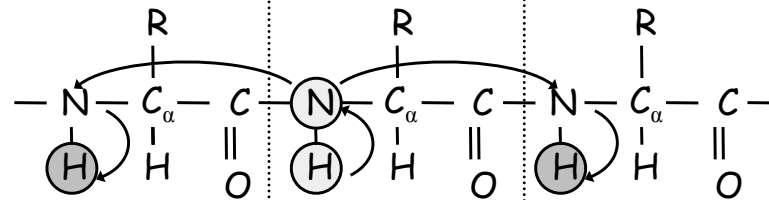
HNCACB /  
HN(CO)CACB



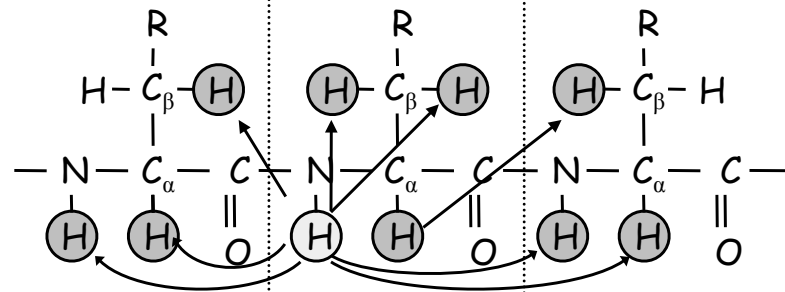
HNCO /  
HN(CA)CO



HN(CACO)NH

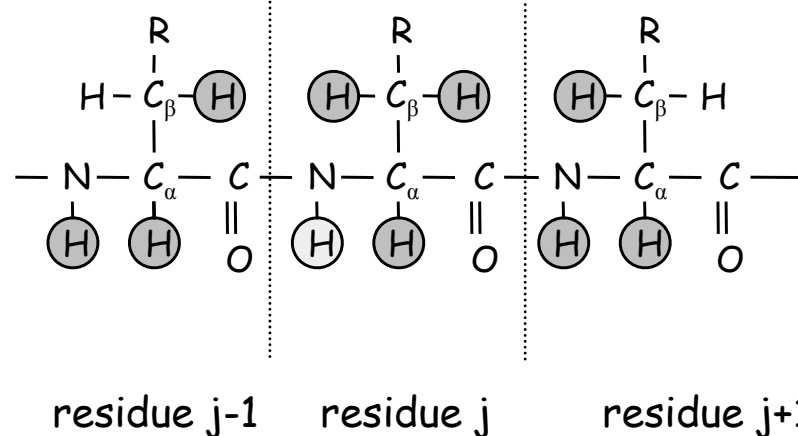
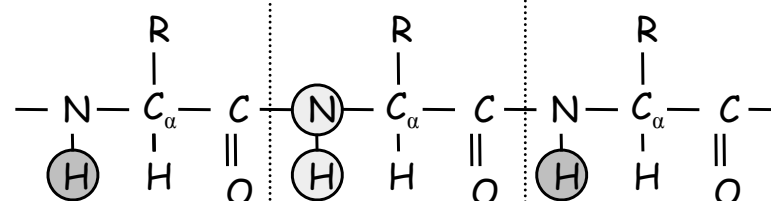
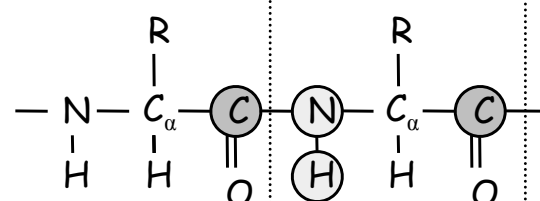
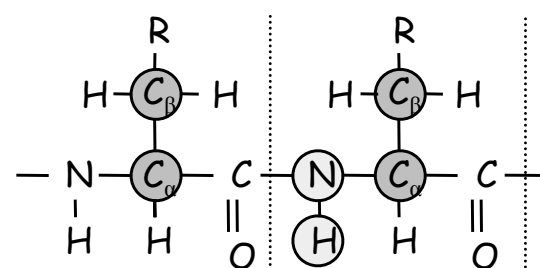
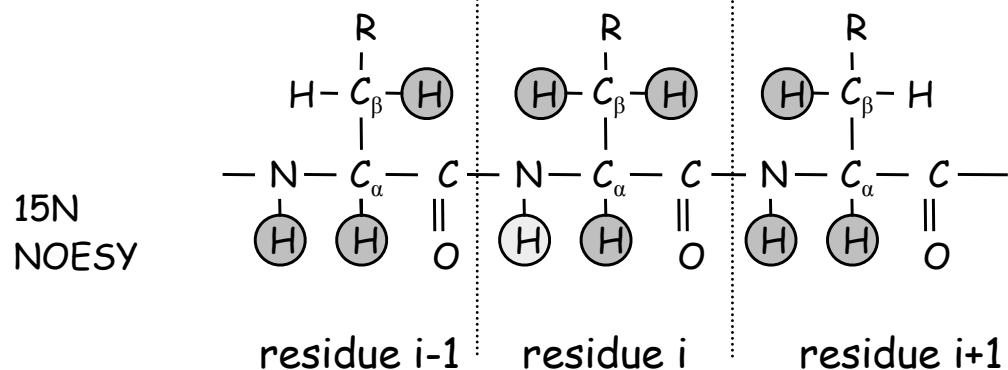
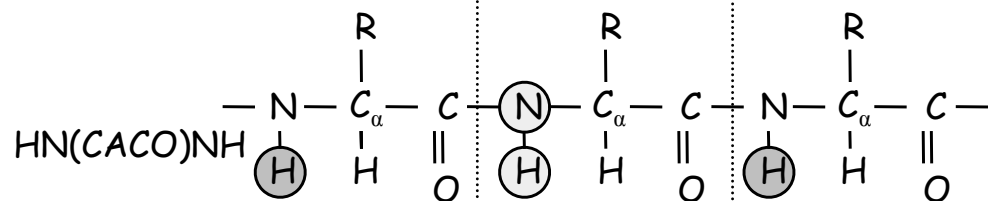
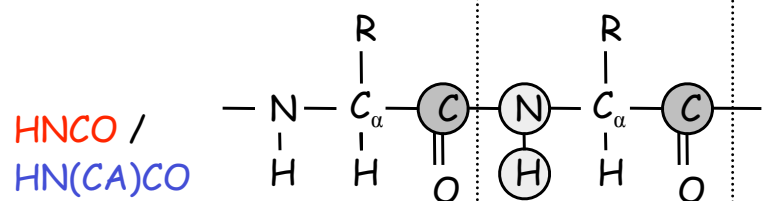
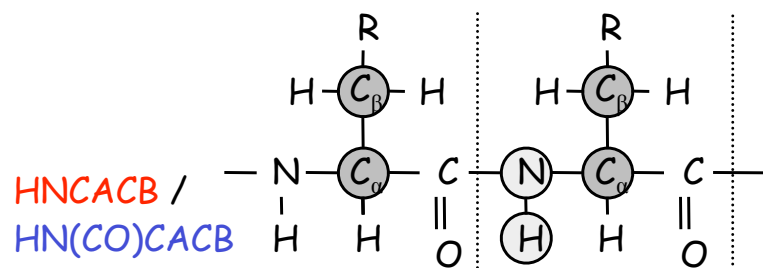


<sup>15</sup>N  
NOESY

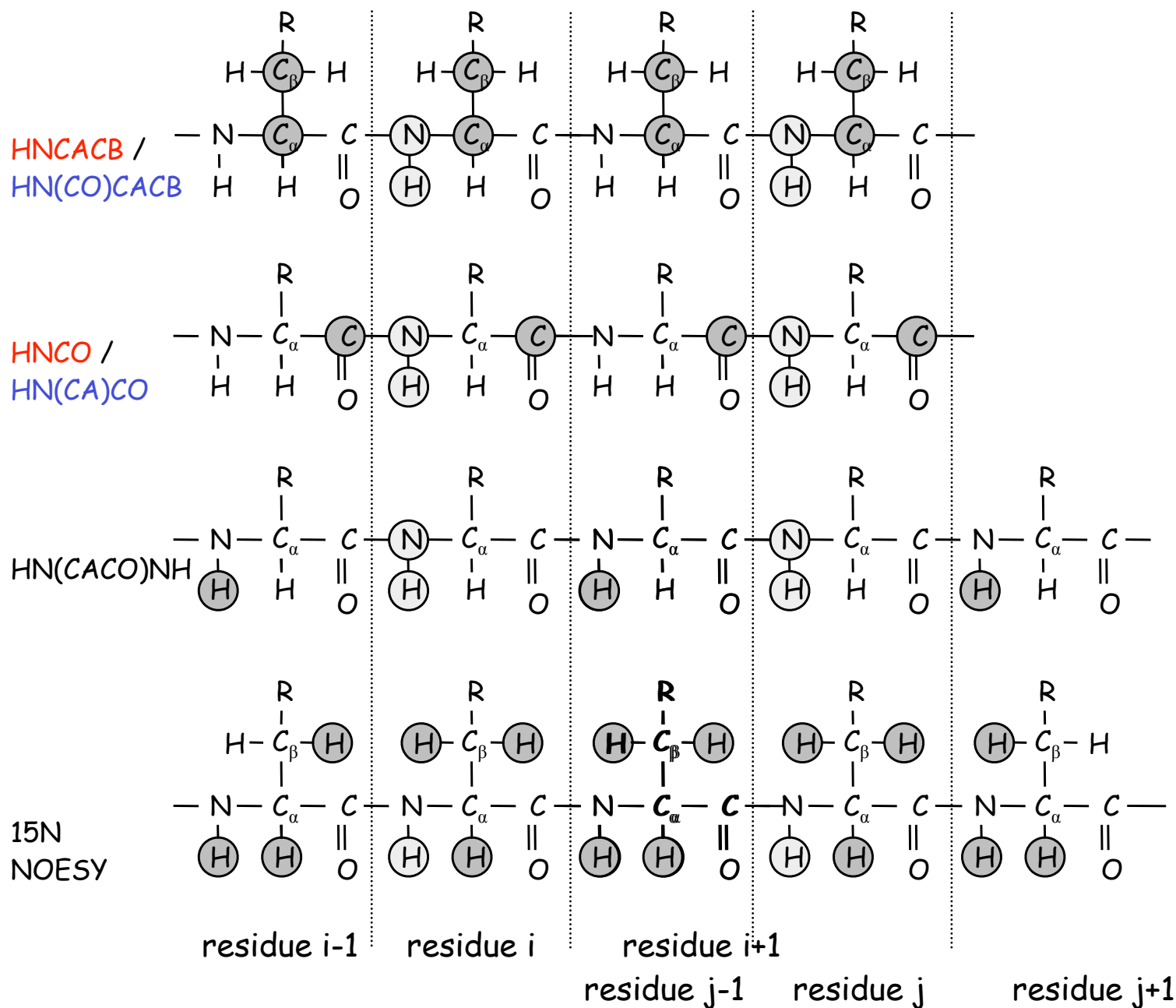


residue i-1    residue i    residue i+1

# Backbone Assignment - 3D Experiments

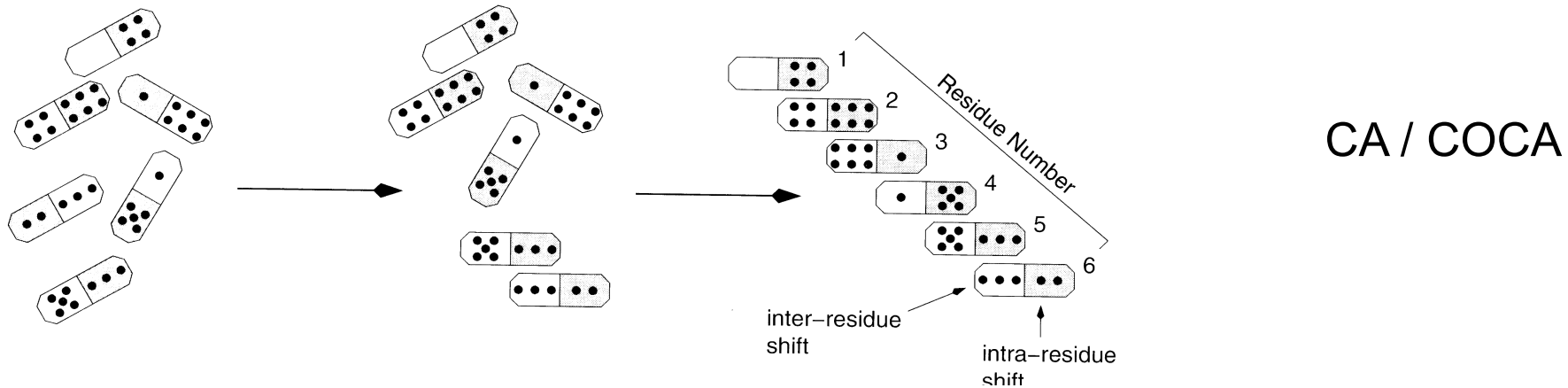


# Backbone Assignment - 3D Experiments



# sequential assignment strategy: building of fragments

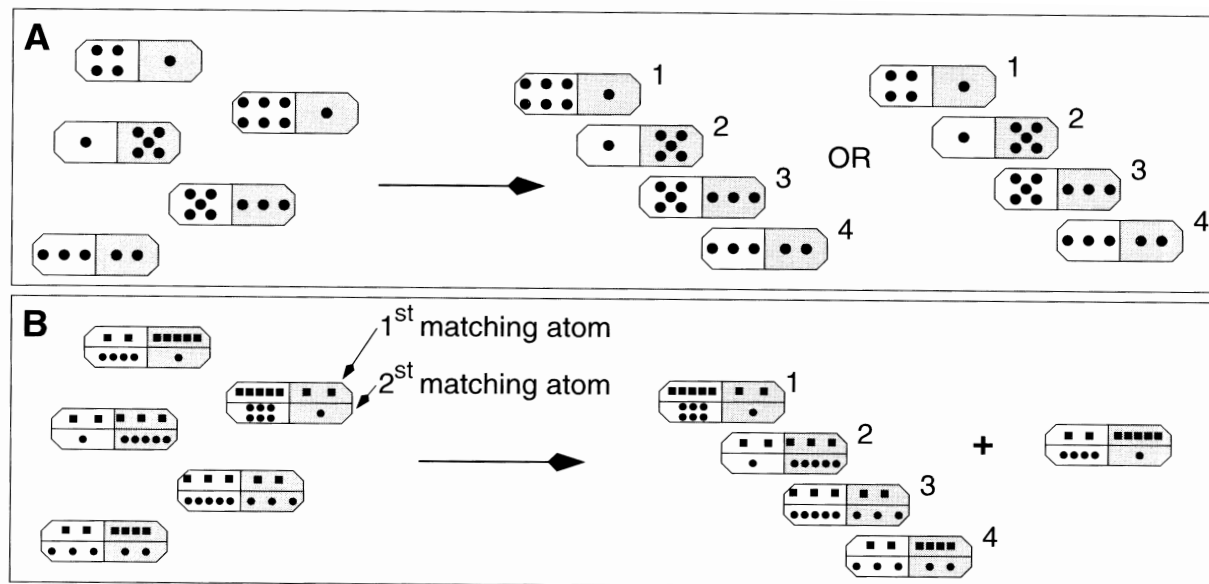
1. picking of HN-C peaks -> spin systems (numbers 1-125)
2. alignment of  $^{13}\text{C}$  dimension for linking of correct successors/ predecessors



CACB / COCACB

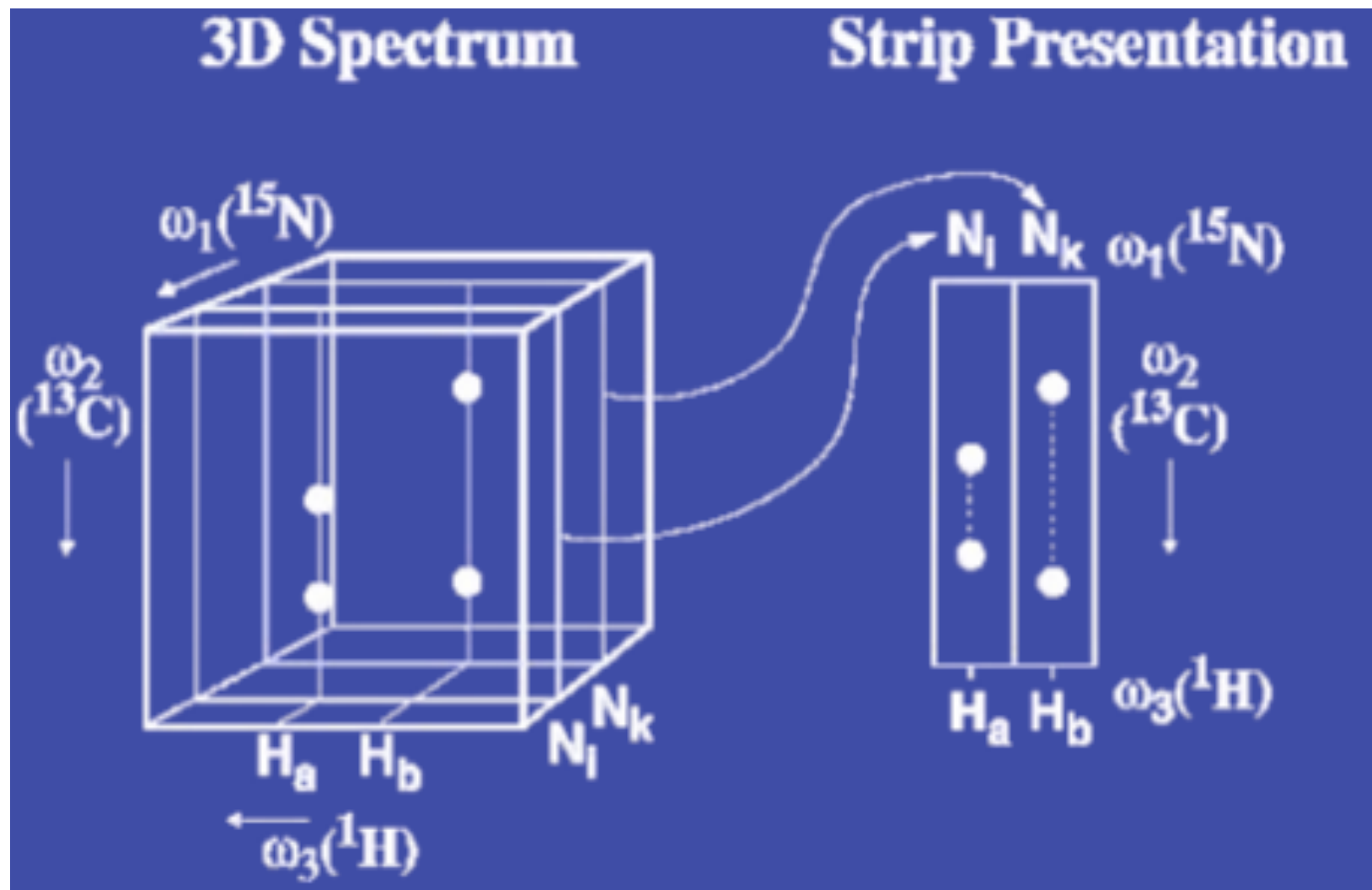
CG / COCG

NN



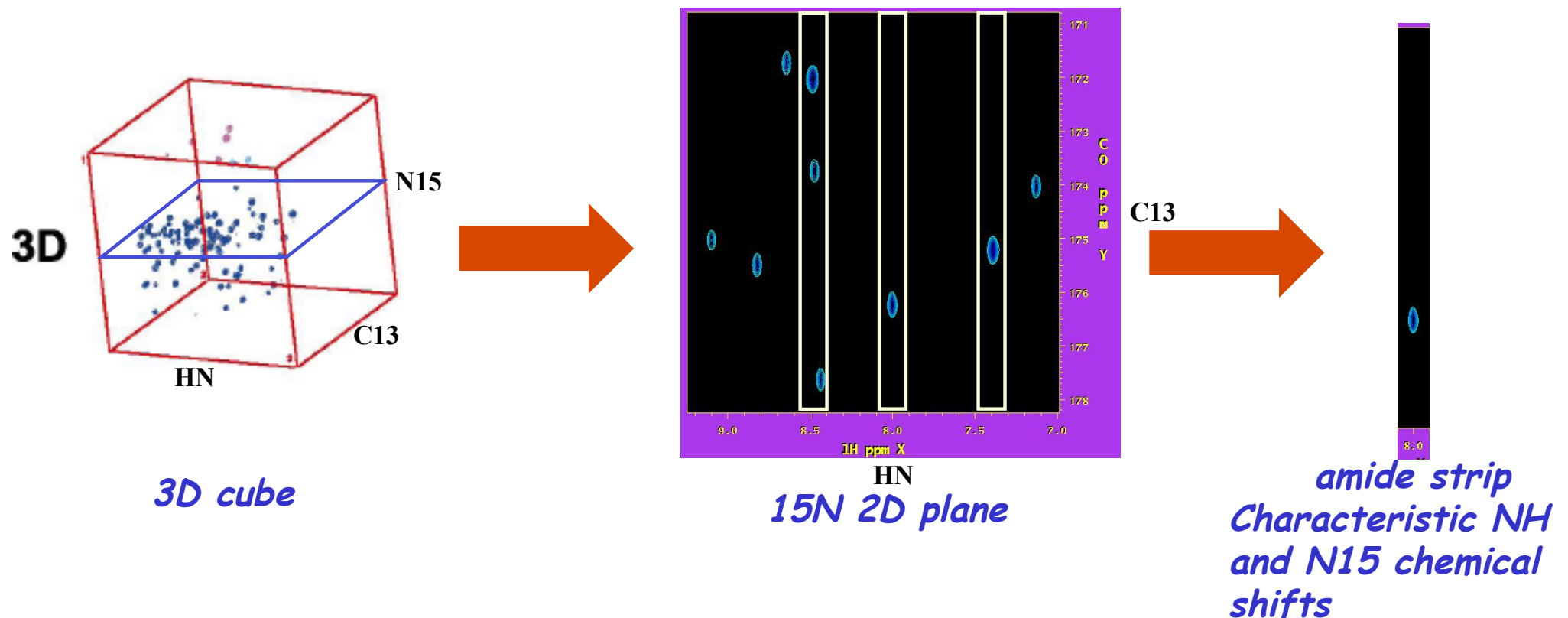
3. try to match fragments on amino acid sequence

# view of the 3D spectrum in CARA



2D  $^1\text{H}$ - $^{15}\text{N}$  HSQC is the root experiment of most of the standard *triple-resonance* ( $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^{15}\text{N}$ ) NMR experiments used for backbone assignment.

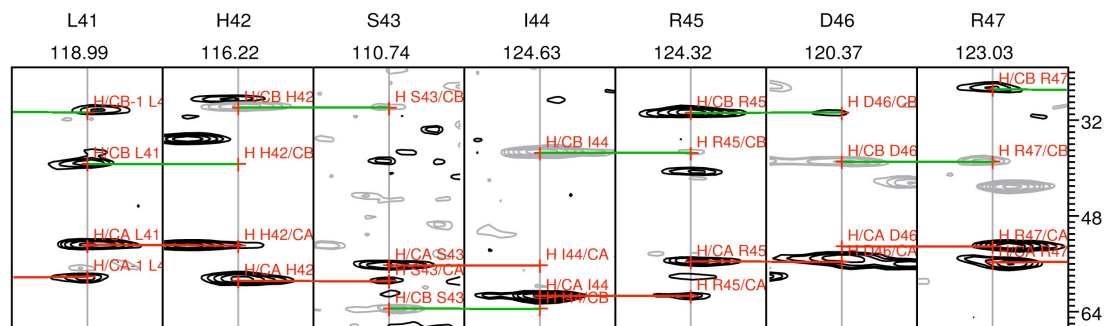
All the 3D triple resonance experiments are related by the common  $^1\text{H}$ ,  $^{15}\text{N}$  chemical shifts of the HSQC spectra: **AMIDE STRIP**



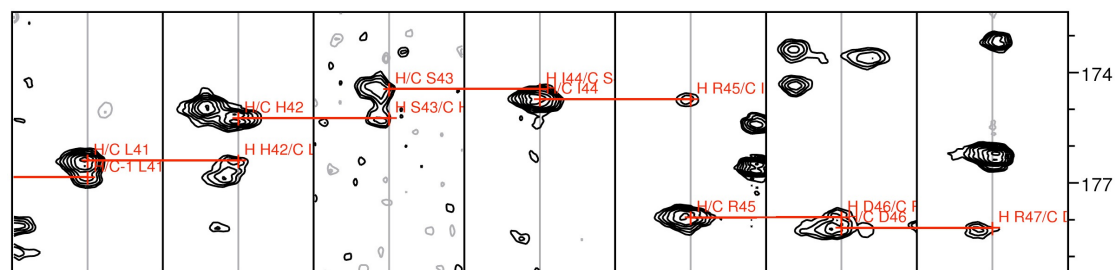


# Backbone Assignment - 3D Experiments

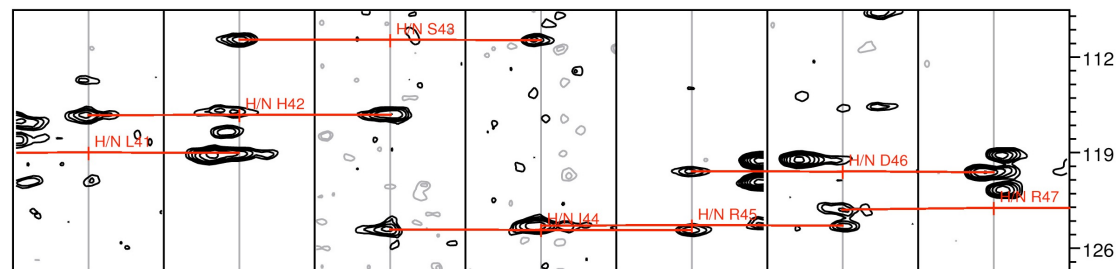
$^1\text{H}/^1\text{H}$  CACB /  
 $^1\text{H}/^1\text{H}$  (CO) CACB



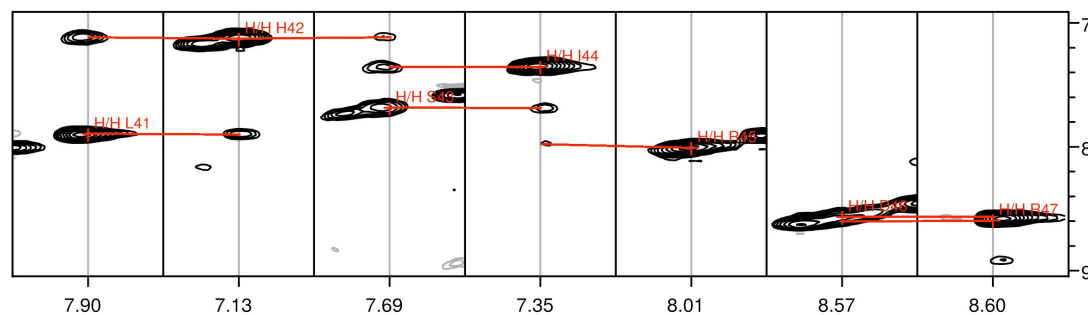
$^1\text{H}/^1\text{H}$  CO /  
 $^1\text{H}/^1\text{H}$  (CA) CO



$^1\text{H}/^1\text{H}$  (CA) CO /  $^1\text{H}/^1\text{H}$  (CA) CO



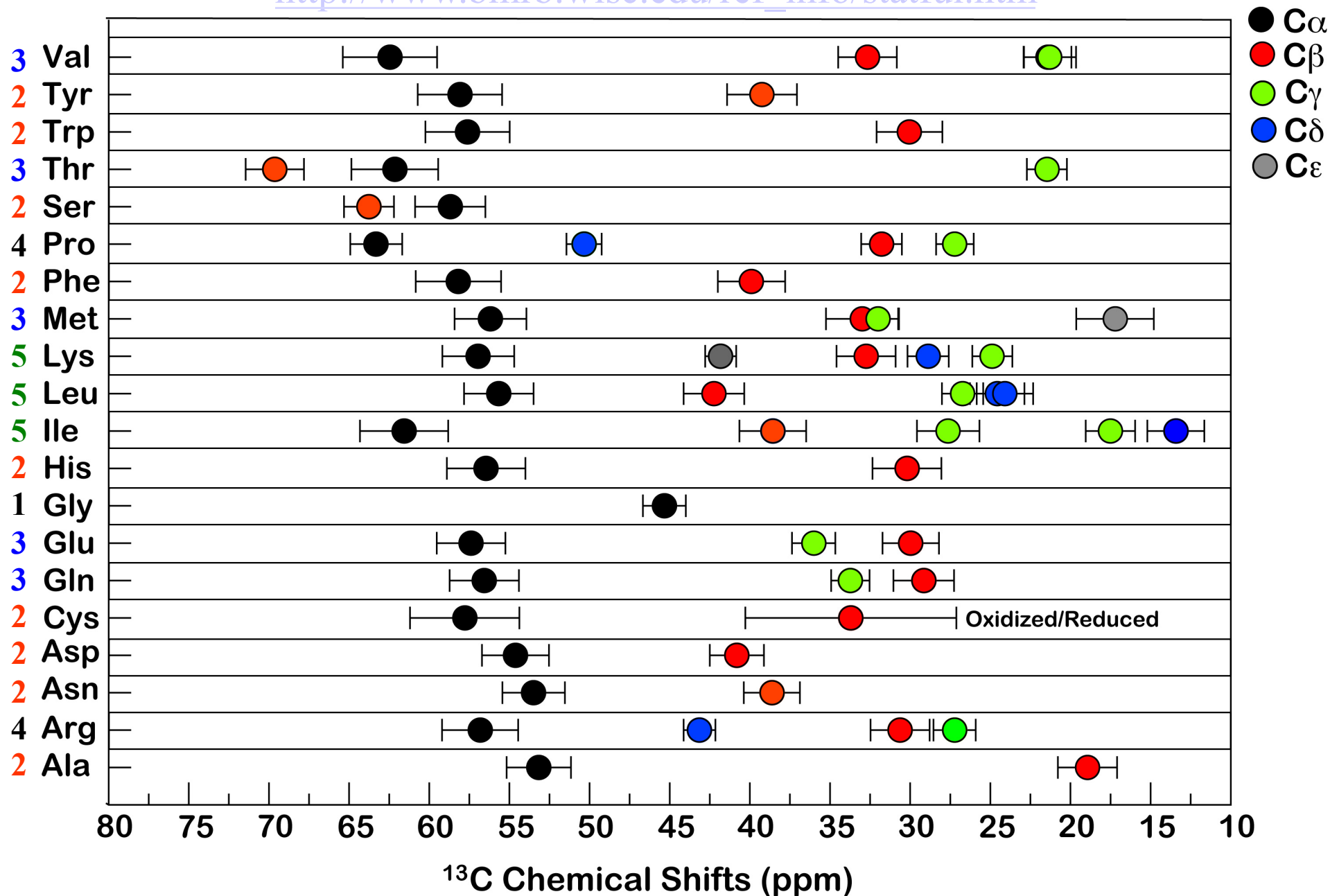
$^{15}\text{N}$   
NOESY



# Standard Carbon Chemical shifts

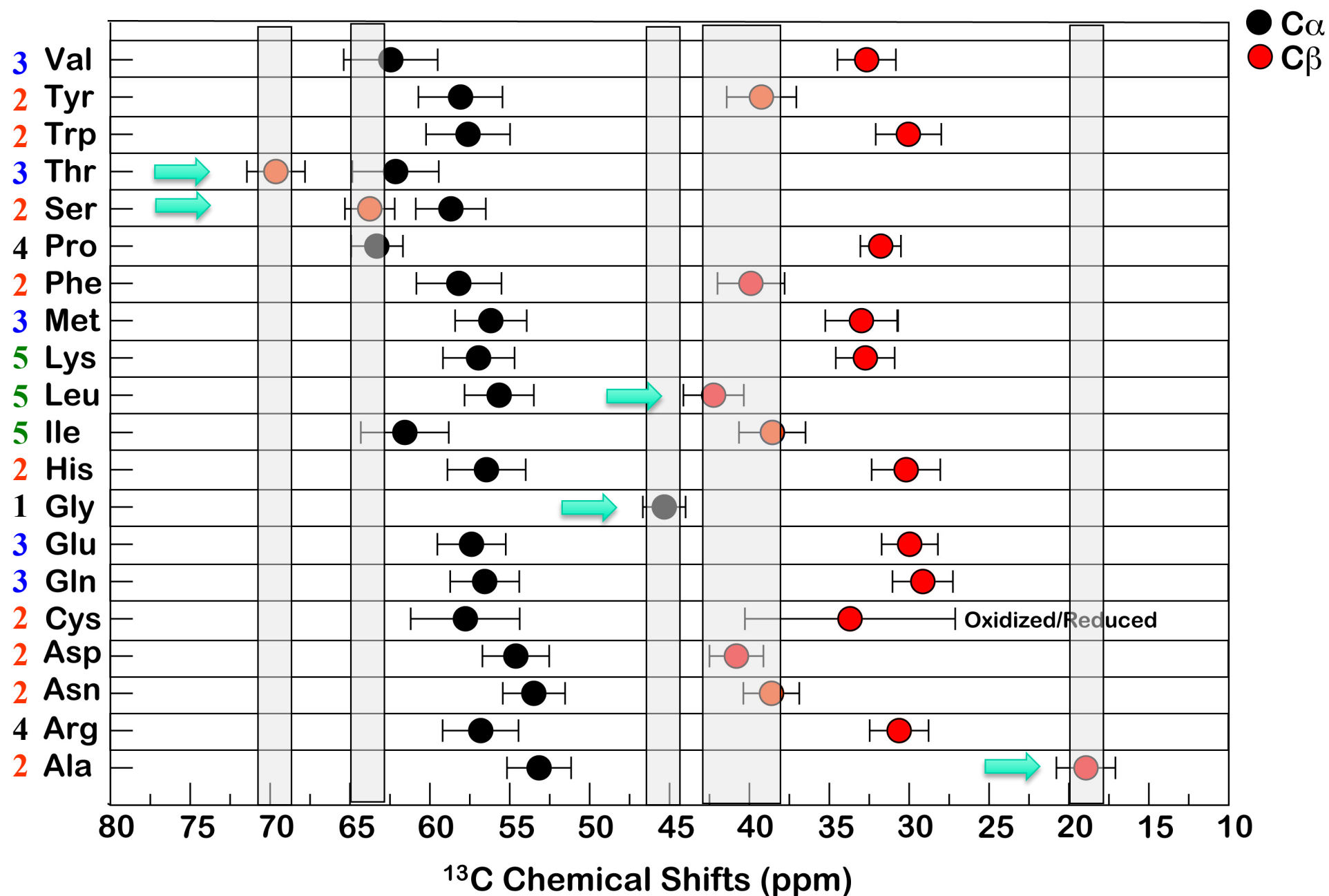
Biological Magnetic Resonance Databank (Diamagnetic Shifts - 03/09/2007)

[http://www.bmrb.wisc.edu/ref\\_info/statful.htm](http://www.bmrb.wisc.edu/ref_info/statful.htm)



# Standard Carbon Chemical shifts

Biological Magnetic Resonance Databank (Diamagnetic Shifts - 03/09/2007)



# Identifying Residue type from chemical shifts

Can easily identify Gly, Ser/Thr and Ala from  $C_{\alpha}$ ,  $C_{\beta}$  shifts: Gly  $C_{\alpha}$  ~45 ppm; Ser and Thr can be distinguished by  $C_{\beta}$  shifts: Thr  $C_{\beta}$  ~70 ppm ; Ser  $C_{\beta}$  ~ 63 ppm. Ala  $C_{\beta}$  chemical shifts is around ~18 ppm.

Can group Leu, Tyr, Phe, Asn, Ile and Asp based on their  $C_{\beta}$  shifts ~> 35 ppm.

Differentiate between the residues having two (Asp, Asn, Trp, Tyr, Cys, His, Phe) carbons sidechains and those having 3 or more carbons in the sidechain by using  $CC(CO)NH$ .

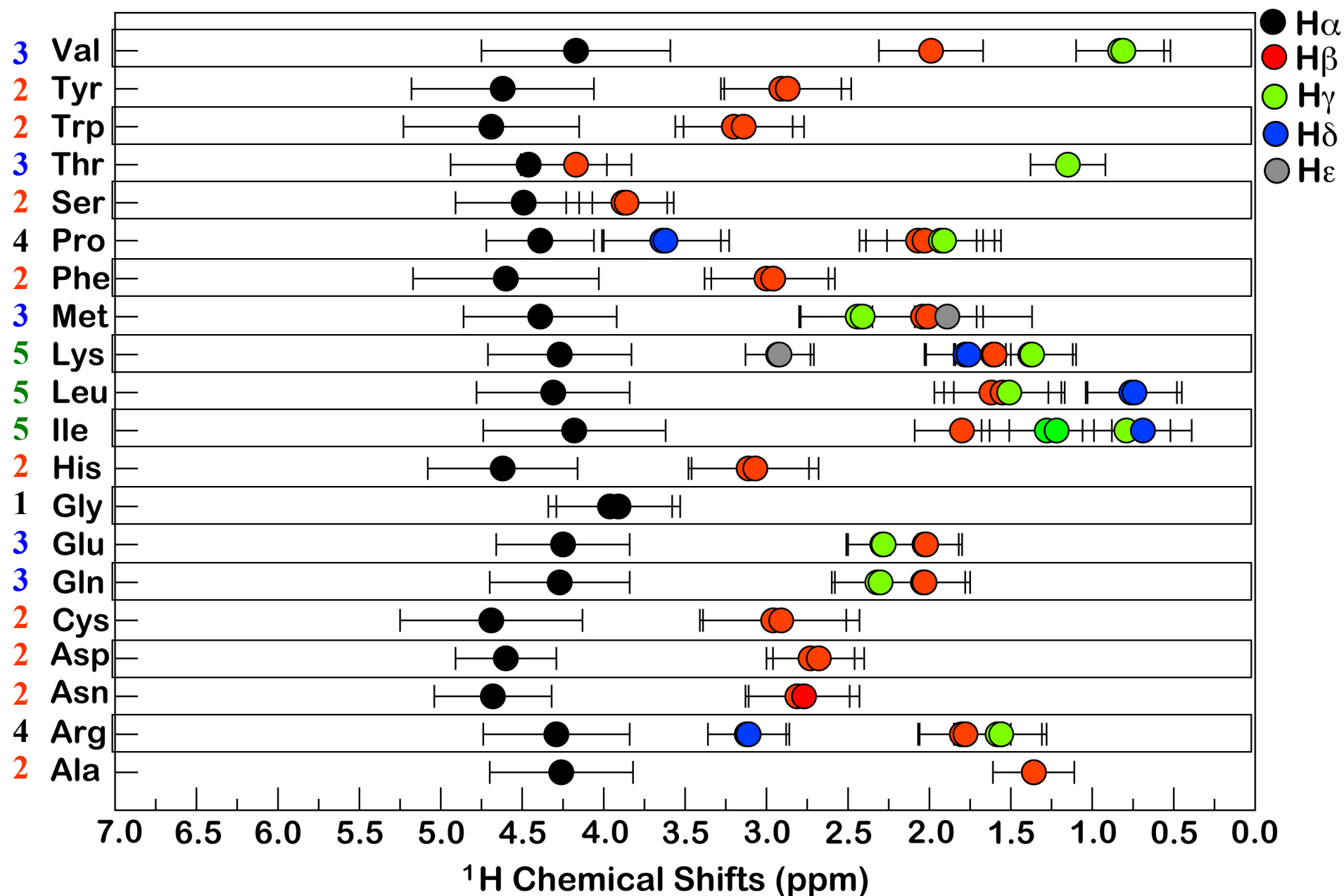
Among the residues having three carbon sidechain: Val, Met, Thr, Glu and Gln, Val has most upfield  $C_{\gamma}$  chemical shifts. Ser/Thr can be distinguished  $C_{\gamma}$  shift. Glu and Gln can be identified by their  $C_{\gamma}$  shifts, Glu  $C_{\gamma}$  > 35 ppm and Gln  $C_{\gamma}$  < 35 ppm.

Residues with four carbons sidechain: Pro and Arg can easily be distinguished by their  $C_{\delta}$  shifts, for Pro  $C_{\delta}$  ~ 50 ppm whereas for Arg  $C_{\delta}$  ~43 ppm.

Among the residues having five carbons side chain: Leu, Lys and Ile. Ile has the most upfield  $C_{\delta}$  shifts ~10 ppm whereas Leu has  $C_{\beta}$  ~ 43 ppm and Lys will have  $C_{\epsilon}$  ~43 ppm.

# Standard Proton Chemical shifts

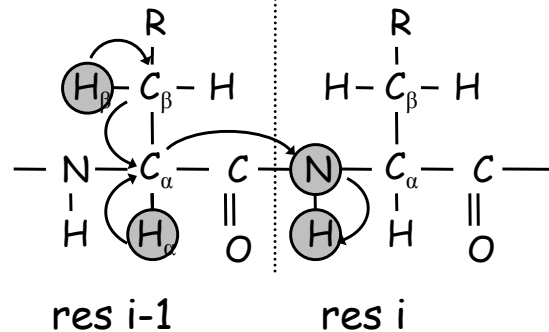
Biological Magnetic Resonance Databank (Diamagnetic Shifts - 03/09/2007)



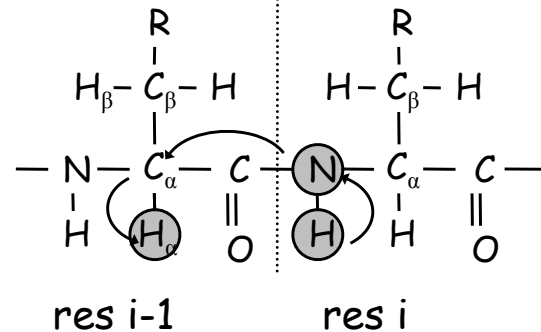
# Side Chain Assignment Strategies

Identification of backbone protons:

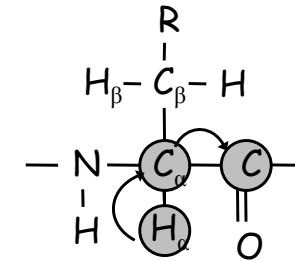
HBHA(CACBCO)NH



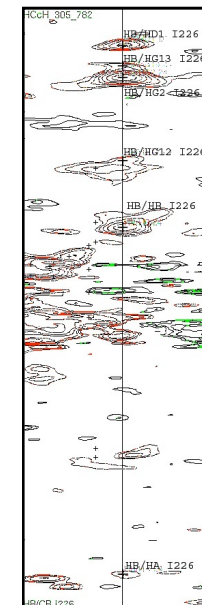
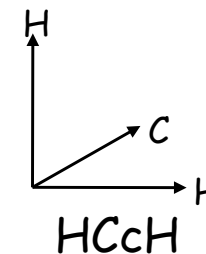
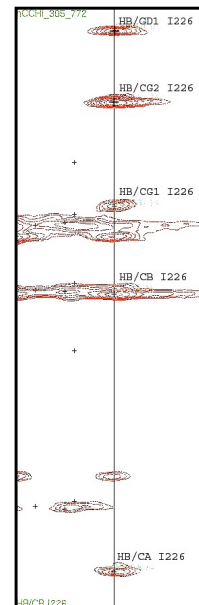
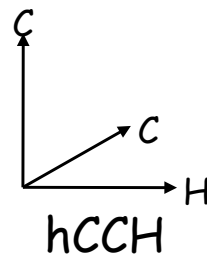
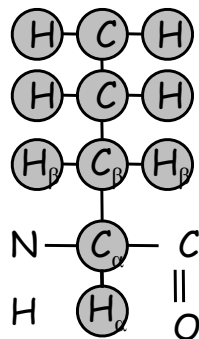
HN(COCA)HA



HACACO

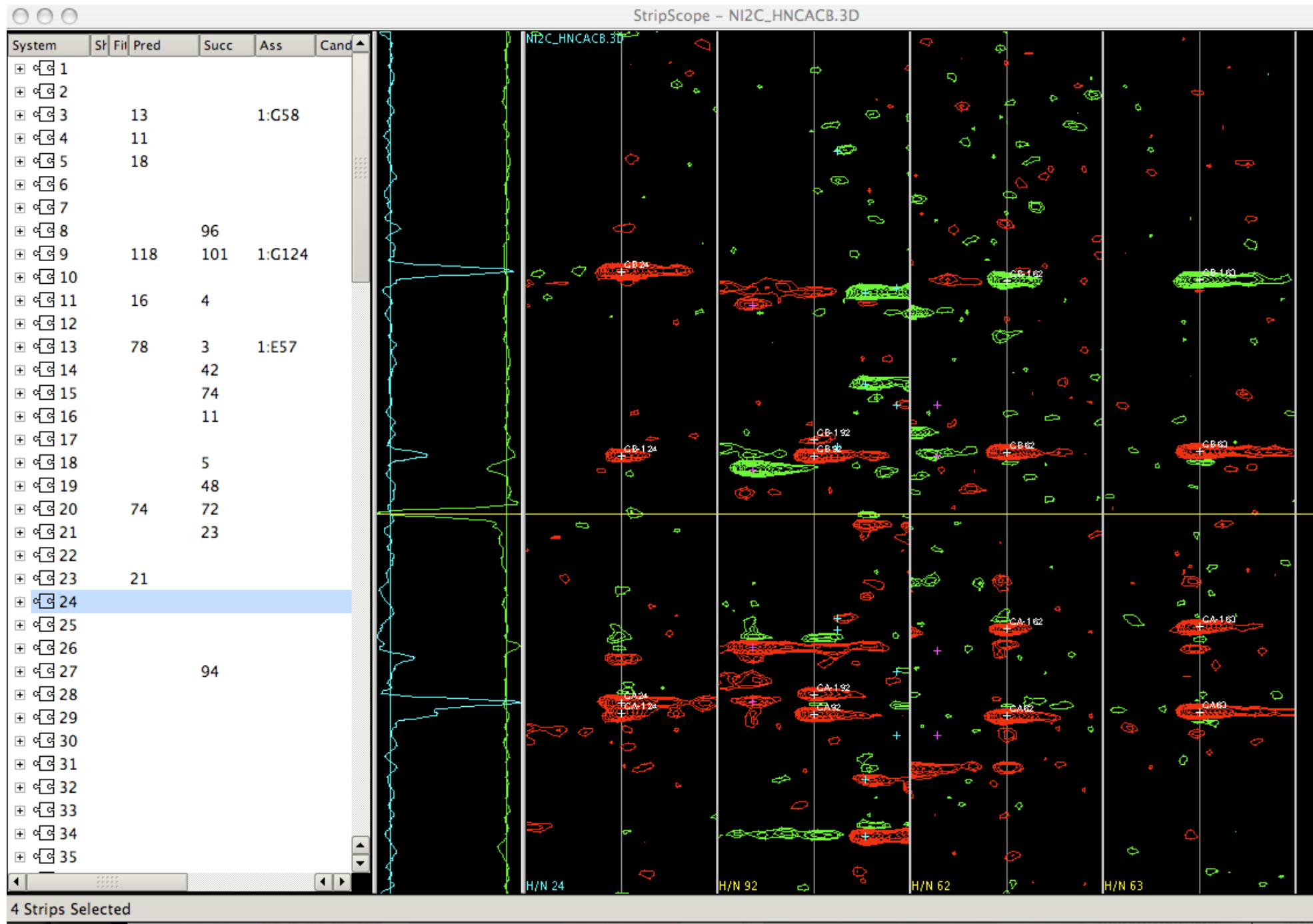


Side chain assignment:

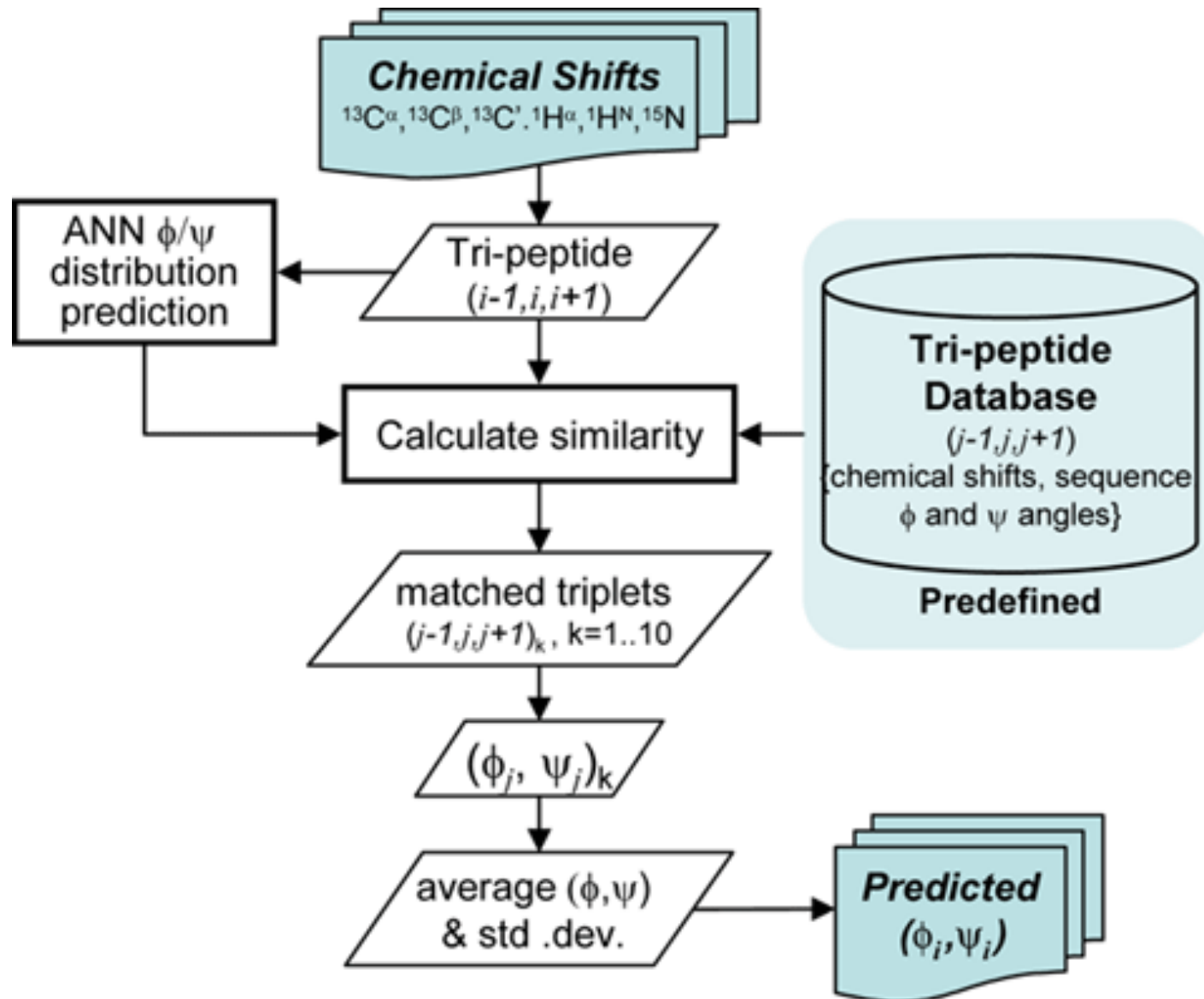




# $^{13}\text{C}$ dimension strips of single spin systems for linking



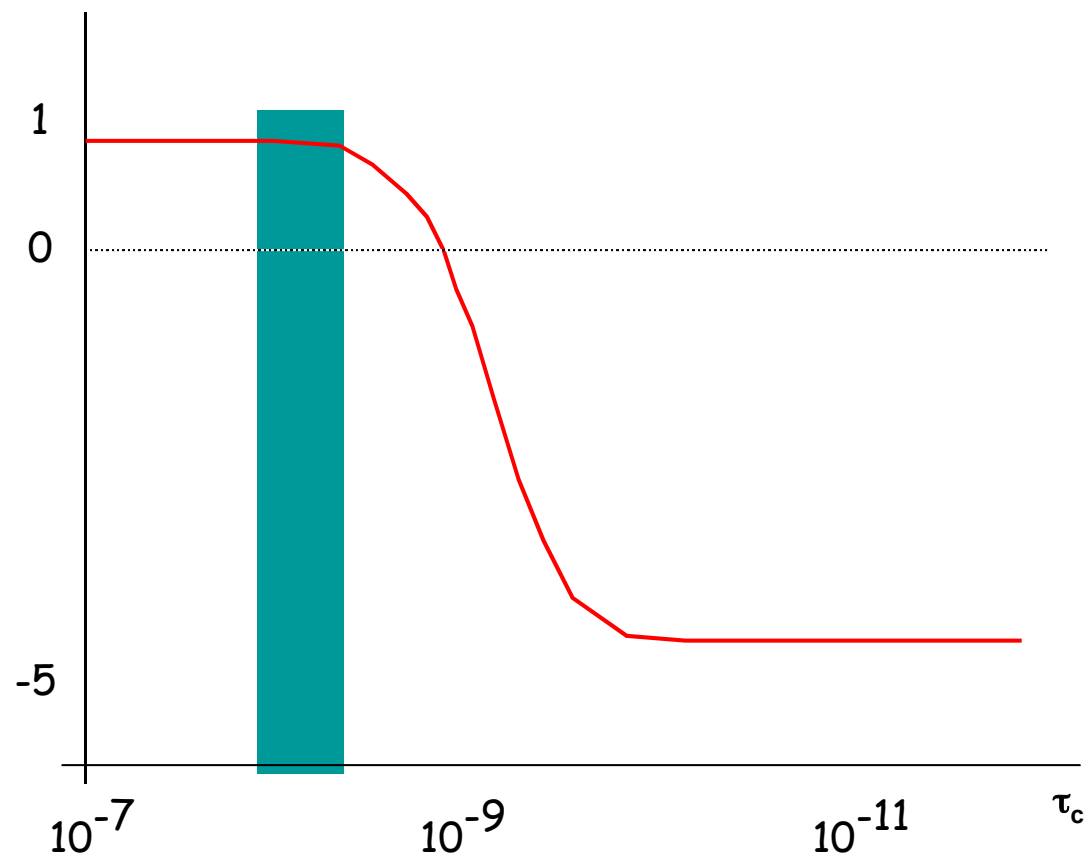
# Prediction of secondary structure using chemical shifts using TALOS



# The magnitude of the $^1\text{H}\{^{15}\text{N}\}$ -NOE depends on the motional properties

$^{15}\text{N}\{^1\text{H}\}$

-NOE

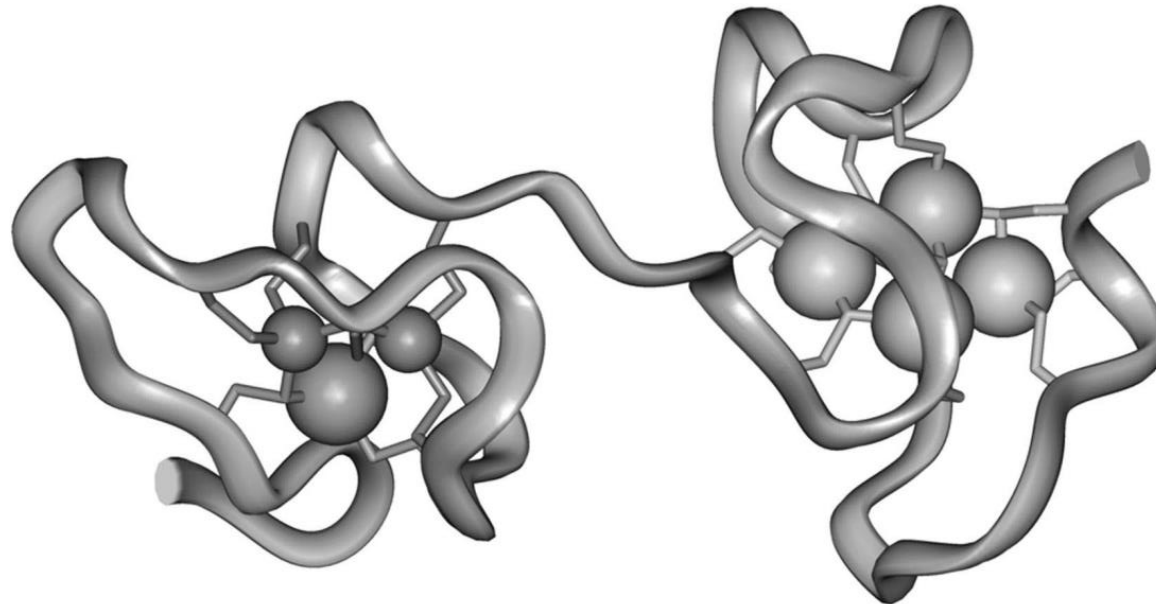


Correlation time

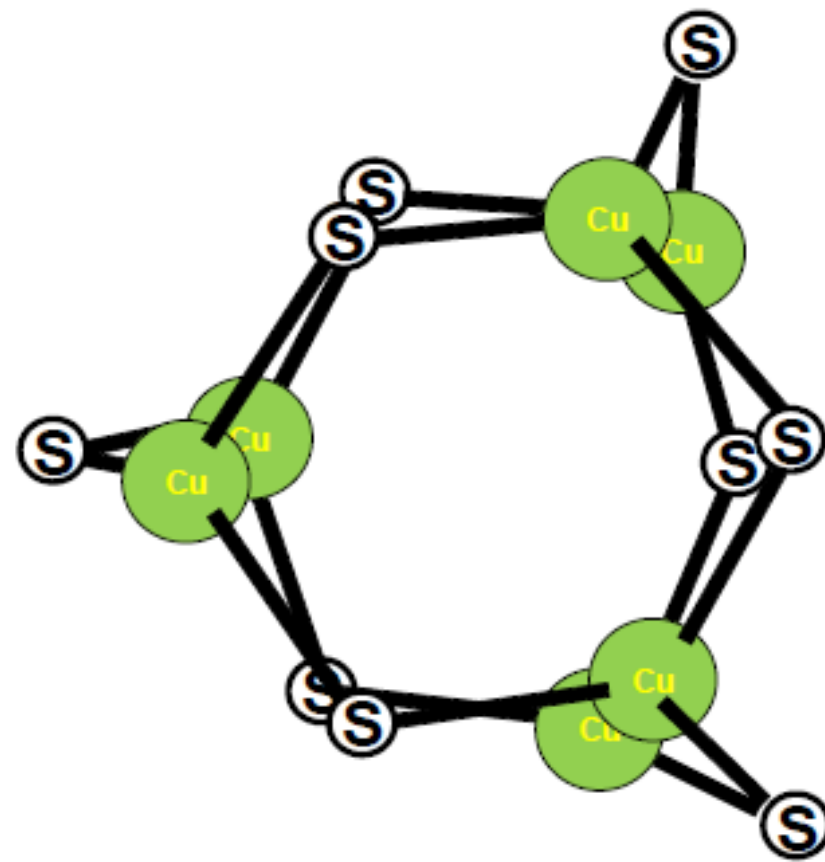
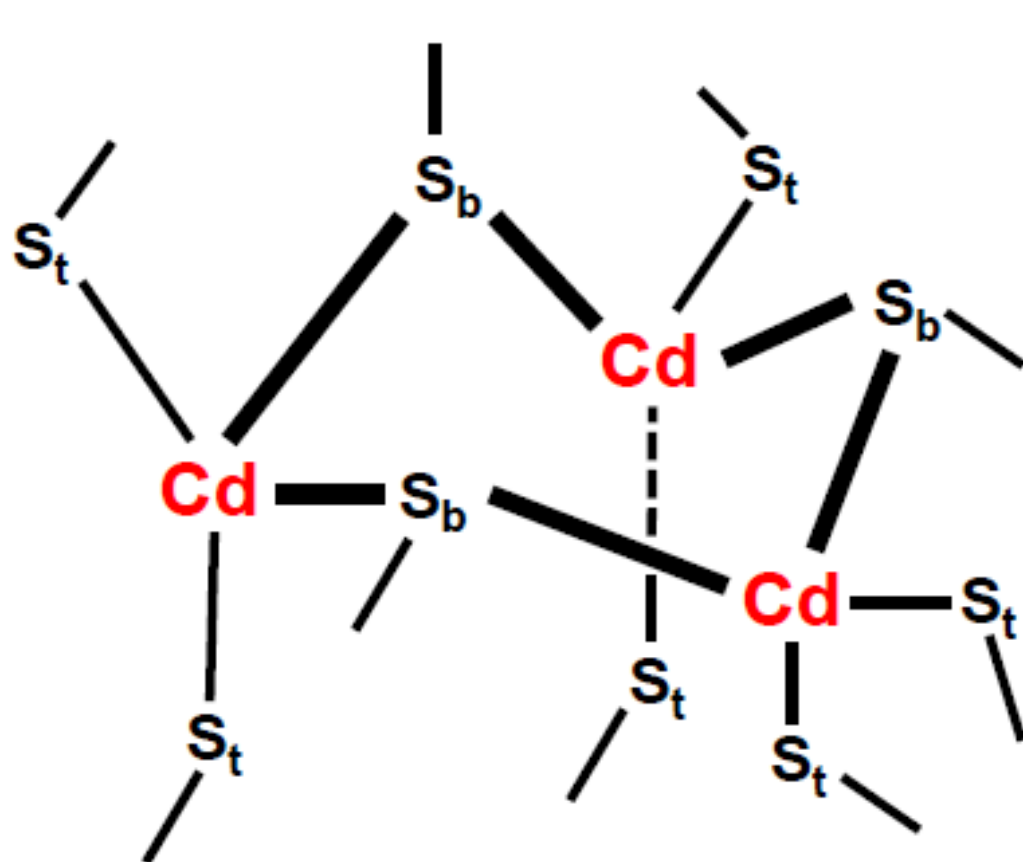
# NMR of metallothioneins

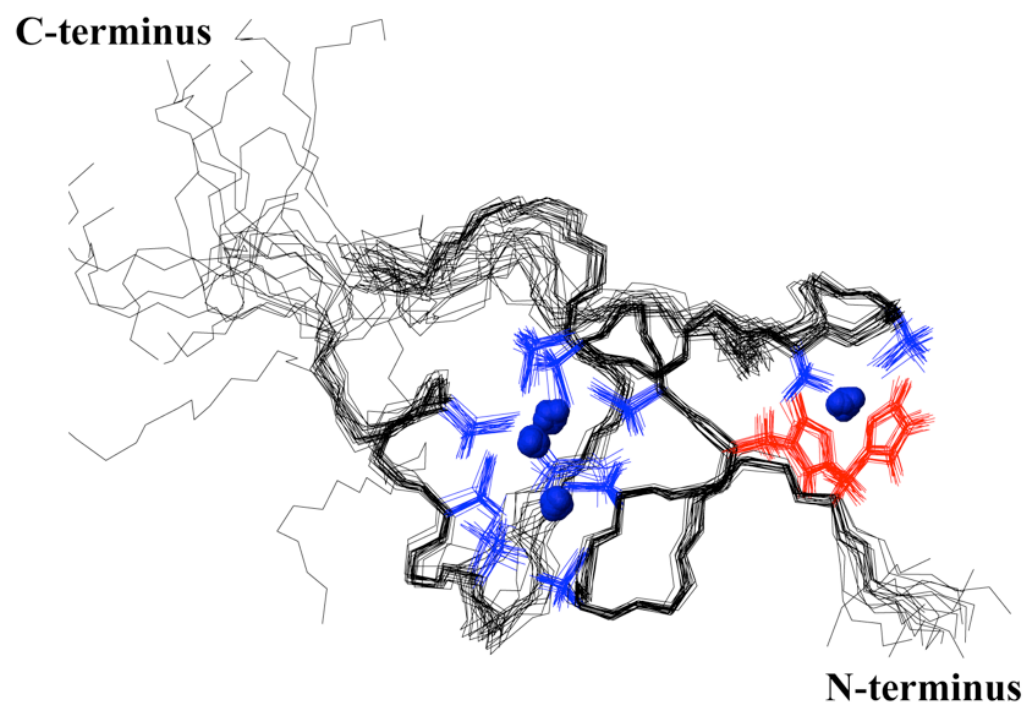
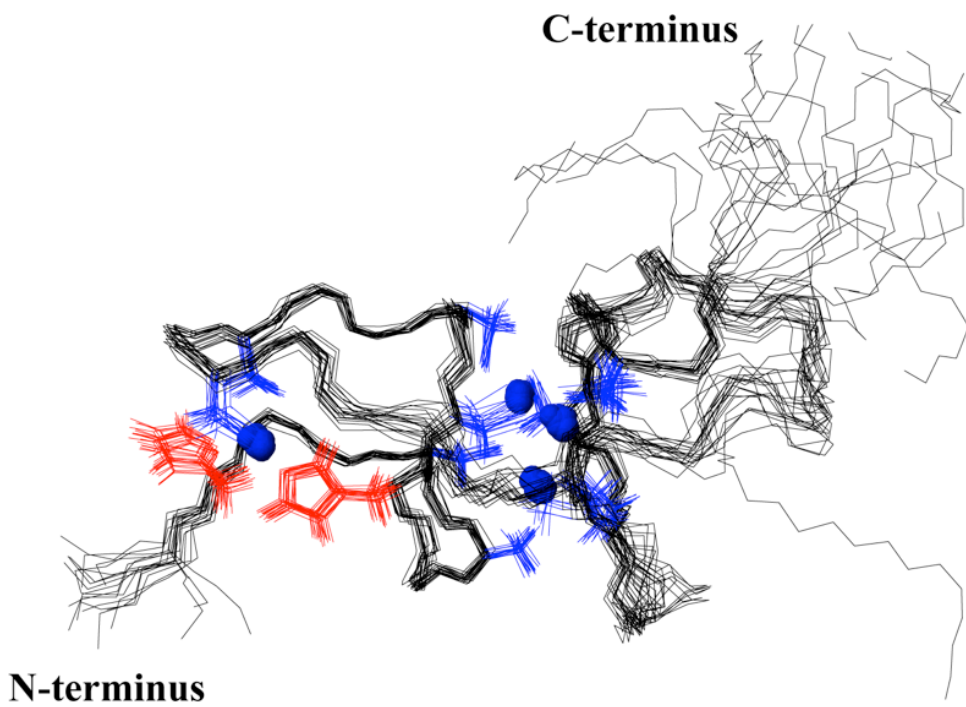
# Metallothioneins

- Small proteins: ~60 aa with ~30% cysteine residues
- Coordinate metal ions
- No secondary structure elements
- Two metal-thiolate clusters per protein:
  - $\alpha$ -domain: 11 Cys coordinating 4 divalent metal ions
  - $\beta$ -domain: 9 Cys coordinating 3 divalent metal ions

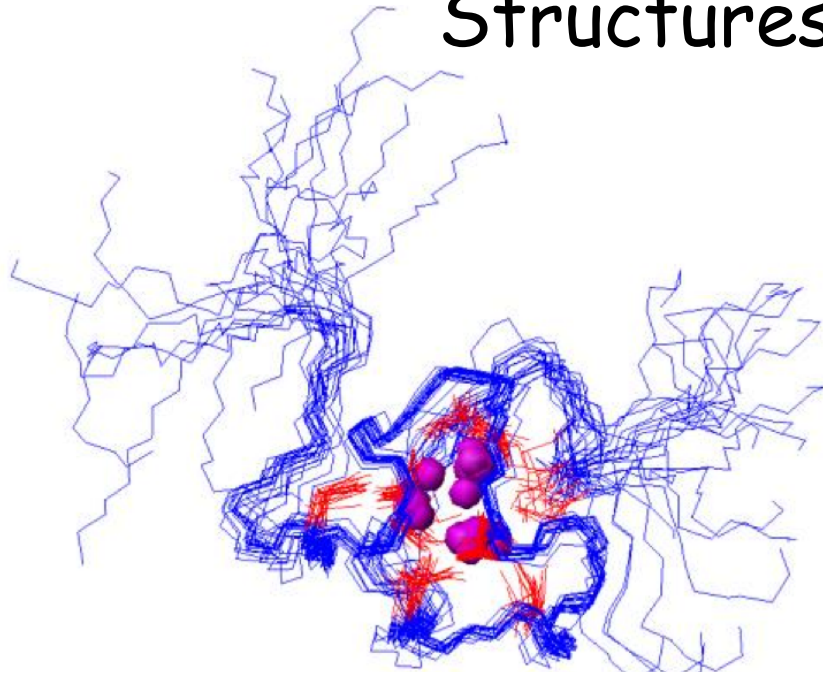


B

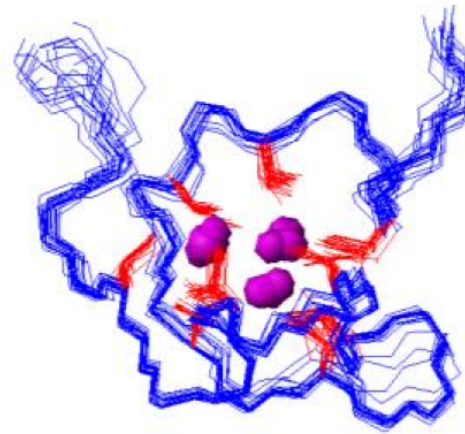




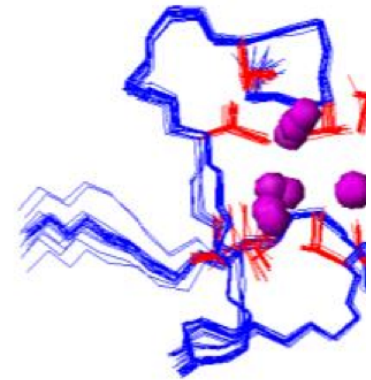
# Structures of *Littorina littorea* MT



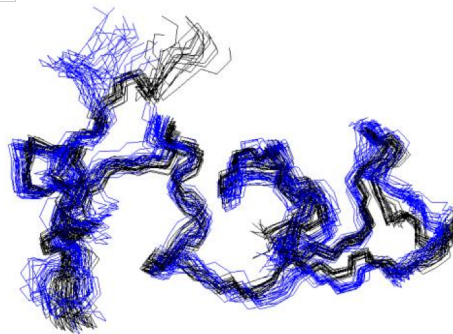
**N-term**



**center**



**C-term**



**center and C-term**